

PC-3 Cells | 300312

Description

PC3 cells, derived from the bone metastasis in a 62-year-old Caucasian male with grade IV prostatic adenocarcinoma, are a cornerstone in the study of human prostate carcinoma. The PC-3 human prostate cancer cell line is widely used for studying the molecular and cellular aspects of prostate cancer, especially in the context of metastatic disease. Their high metastatic potential makes them a valuable model for advanced prostate cancer research.

As epithelial cells, PC3 cells' lack of response to androgens and their independence from typical growth factors like glucocorticoids or fibroblast growth factors, positions them uniquely among human prostate carcinoma cells for studying the impact of koenimbin and other potential therapeutic agents.

The absence of prostate-specific antigen (PSA) expression and low activities of testosterone-5-alpha reductase and acidic phosphatase set PC3 apart from other prostate cancer cell models like LNCaP and DU145, the former known for expressing luminal differentiation markers such as AR and PSA, and the latter representing a moderated metastatic potential of prostate carcinoma.

Furthermore, the role of the PC3 prostatic carcinoma cell line in prostate cancer stem cells research is underscored by the observation that a subset forms cancer stem cell holoclones. This characteristic makes the PC3 cell line a critical model for studying the tumor environment, particularly through xenograft models where PC3 xenograft tumors are used to investigate tumor growth and response to therapies in vivo.

In summary, PC3 cells, originating from a grade IV prostatic adenocarcinoma, serve as a pivotal model in prostate cancer research due to their high metastatic potential, unique androgen independence, and distinct cellular characteristics. Their versatility extends from molecular studies of metastasis to the exploration of therapeutic responses and the investigation of prostate cancer stem cells, making them an invaluable resource for advancing our understanding of prostate carcinoma's complexities and potential treatments.

Organism Human

Tissue Prostate

Disease Adenocarcinoma

Metastatic site Bone

Applications Transfection host

Synonyms PC-3, PC.3

Age 62 years

Gender Male

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Ethnicity Caucasian

Morphology Epithelial-like

Growth properties Adherent. The cells form clusters in soft agar and can be adapted to suspension growth

Citation PC3 (Cytion catalog number 300312)

Biosafety level 1

NCBI_TaxID 9606

CellosaurusAccession CVCL_0035

Antigen expression HLA A1, A9

Tumorigenic Yes, in nude mice

Karyotype The karyotype of PC3 cells is notable for being triploid, containing multiple chromosomal abnormalities that contribute to their aggressive nature.

Culture Medium DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO₃ (Cytion article number 820400a)

Supplements Supplement the medium with 5% FBS

Dissociation Reagent Accutase

Doubling time 40 hours

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Subculturing Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Seeding density Start with 3×10^4 cells/cm². After cell recovery, use the seeding density of 1×10^4 cells/cm² for the subsequent splitting steps.

Fluid renewal 2 to 3 times per week

Post-Thaw Recovery After thawing, plate the cells at 5×10^4 cells/cm² and allow the cells to recover from the freezing process and to adhere for at least 24 hours.

Freeze medium As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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**Thawing and
Culturing Cells**

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

**Incubation
Atmosphere**

37°C, 5% CO₂, humidified atmosphere.

**Shipping
Conditions**

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

**Storage
Conditions**

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.