Product sheet

Cytion

B95-8 Cells | 601102

Description	The B95-8 cell line is an immortalized marmoset B lymphoblastoid line, derived from the peripheral blood leukocytes of a cotton-top marmoset (Saguinus oedipus). This cell line was established through infection with the Epstein-Barr virus (EBV), which is a common method for immortalizing B cells. The presence of EBV is central to the B95-8 line's utility in research, particularly for studies related to viral oncology, virus-host interactions, and the biology of EBV itself. B95-8 cells are frequently used as a source of Epstein-Barr virus in virology research. They produce infectious virus particles, making them an invaluable tool for the propagation of EBV and for experiments requiring active virus. Additionally, this cell line has been instrumental in the development of vaccines and therapeutic strategies against EBV-associated diseases, including Burkitt's lymphoma and Hodgkin's lymphoma. The cells are also relevant in the study of the immune response to EBV, as they can be used to model the transformation of B cells and to understand the mechanisms of EBV-induced tumorigenesis.
Organism	Cotton-top tamarin
Tissue	Blood
Synonyms	B95.8, B 95.8, B 95-8, B-95-8, B958, GM07404, GM07404A, GM07404D

Gender	Female
Morphology	Lymphoblast
Growth properties	Suspension

Citation	B95-8 (Cytion catalog number 601102)
Biosafety level	2
NCBI_TaxID	9490
CellosaurusAccession	CVCL_1953

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Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with 10% FBS
Subculturing	Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x 10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.

2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.

3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.

4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.

5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.

6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.

7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.

8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

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Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.