Product sheet

HS-695T Cells | 300211

Description	The HS-695T cell line is derived from human melanoma, a type of skin cancer characterized by the malignant transformation of melanocytes. These cells were originally obtained from an adult patient and have since been extensively utilized in research focused on melanoma biology, tumorigenesis, and cancer metastasis. The HS-695T cell line exhibits key characteristics of melanoma, including the ability to proliferate rapidly and form tumors when transplanted into immunocompromised mice. This cell line retains many of the molecular and genetic features of the original tumor, making it a valuable model for studying the underlying mechanisms of melanoma progression and for testing potential therapeutic agents. HS-695T cells express various melanoma-associated markers, including Melan-A, tyrosinase, and HMB-45, which are commonly used to identify and study melanocytic tumors. These cells are also known to have mutations in genes such as BRAF and NRAS, which are frequently observed in melanoma and contribute to the oncogenic signaling pathways driving tumor growth and survival. Researchers use the HS-695T cell line to explore the effects of targeted therapies, including BRAF and MEK inhibitors, and to investigate the development of resistance to these treatments. Overall, the HS-695T cell line is a critical tool in melanoma research, aiding in the discovery of new therapeutic strategies and improving our understanding of this aggressive cancer.
Organism	Human
Tissue	Skin
Disease	Amelanotic melanoma
Metastatic site	Lymph node
Synonyms	Hs 695.T, Hs-695-T, Hs 695T, HS 695T, Hs695T, HS695T, Hs695

Age	26 years
Gender	Male
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Adherent

Product sheet

HS-695T Cells | 300211

Citation	HS-695T (Cytion catalog number 300211)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0851

Protein expression	P53 positive
Isoenzymes	G6PD, B, PGM1, 1, PGM3, 1, ES-D, 1, Me-2, 0, AK-1, 1, GLO-1, 1, Phenotype Frequency Product: 0.0427
Tumorigenic	Yes, in immunosuppressed mice
Mutational profile	BRAF V600Emut
Karyotype	(P19-40) mode = 52, Y chromosome present

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	2×10^4 cells/cm^2

HS-695T Cells | 300211

Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.
Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.