Product sheet

ARPE-19 Cells | 305025

Description	The ARPE-19 cell line, derived from the retinal pigment epithelium (RPE) of a 19-year-old male, has functional characteristics akin to native RPE cells, making it a pivotal epithelial cell model in ophthalmic research. These cells are utilized in studies related to the vertebrate retina and retinal pigment epithelium physiology. When cultured in 3D cell culture systems or as a cell monolayer on laminin-coated filters with low serum media, ARPE-19 cells achieve morphological polarization and form tight junctions, exhibiting transepithelial resistance akin to that observed in vivo. ARPE-19 cells, expressing RPE-specific markers such as CRALBP and RPE-65, serve as an excellent model for understanding the pigmentation processes of the retinal pigment epithelium, including melanin synthesis and melanosome content. The application of ARPE-19 human cells extends to ocular pharmacokinetics and permeability studies, providing insights into ocular chemotherapy efficacy and retinal barriers considerations. Their use in examining the interactions between pharmacokinetics and melanin content offers valuable data on drug binding and uptake. RPE-19 cells contribute to our understanding of retinal explants and the epithelium's role in eye development, given their expression of networks involved in early eye formation and muscle contraction. In summary, the ARPE-19 cell line serves as a critical model in ophthalmic research, offering insights into the physiology of the retina, pigmentation processes, and the efficacy of ocular treatments.
Organism	Human
Tissue	Eye, retinal pigmented epithelium, retina
Synonyms	ARPE19, Adult Retinal Pigment Epithelial cell line-19, NTC-200, NTC200
Age	19 years
Gender	Male
Morphology	Epithelial
Growth properties	Adherent
Citation	ARPE-19 (Cytion catalog number 305025)

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Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0145

Protein expression	Rpe-Specific Markers Cralbp And Rpe-65
Antigen expression	RPE-specific markers CRALBP and RPE-65
Tumorigenic	Yes

Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO3 (Cytion article number 820400a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO_2, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.