Product sheet

IGROV-1 Cells | 305556

Description	The IGROV-1 cell line is a human ovarian adenocarcinoma cell line extensively utilized in research, particularly in studies involving ovarian cancer. Derived from an ovarian carcinoma, IGROV-1 cells are known for their utility in modeling epithelial ovarian cancer (EOC), which accounts for a majority of ovarian malignancies. This cell line has been employed in various contexts, including evaluating drug responses and mechanisms underlying drug resistance. For example, IGROV-1 has been instrumental in testing the efficacy of targeted therapies, such as the folate receptor alpha-targeting antibody-drug conjugate mirvetuximab soravtansine (IMGN853). This ADC showed promising results by synergizing with chemotherapeutics like carboplatin and doxorubicin, enhancing antitumor efficacy through DNA damage and cell cycle arrest in preclinical models. In addition to its role in cancer research, IGROV-1 has been characterized as a model for viral infection studies. Recent work highlighted its susceptibility to SARS-CoV-2, leveraging its expression of ACE2 to support viral replication. IGROV-1 was shown to mount a robust innate immune response upon infection, similar to primary human nasal epithelial cells, indicating its potential for serological assays, antiviral drug testing, and isolation of viral variants from patient samples. This cell line is considered advantageous for research due to its effective replication of viruses compared to traditional models like Vero cells, which can lead to adaptive mutations. Overall, IGROV-1 cells serve as a valuable model in both oncology and virology, supporting studies of tumor biology, drug resistance, and viral pathogenesis. Their relevance in drug synergy experiments and their compatibility with antiviral research underscore their versatility and importance in the field.
Organism	Human
Tissue	Ovary
Disease	Endometrioid carcinoma
Synonyms	Igrov-1, IGROV 1, IGR-OV1, IGROV1, Igrov1, IGR.OV1, IGROV, OV1/P, OV1/p, OV1-P

Age	47 years
Gender	Female
Ethnicity	Caucasian
Morphology	Epithelial-like
Growth properties	Adherent, monolayer

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Citation	IGROV-1 (Cytion catalog number 305556)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1304

Tumorigenic	Yes, in nude mice.
Mutational profile	Mutation: BRCA1, p.Lys654Serfs*47 (c.1961delA), heterozygous; Mutation: BRCA2, p.Lys1108Argfs*11 (c.3323delA) (p.Gln1107fs) (c.3320delA); Mutation: PIK3CA, p.Arg38Cys (c.112C>T), heterozygous; Mutation: PIK3CA, p.Ter1069TrpinsLysAspAsn (c.3207A>G), heterozygous; Mutation: PTEN, p.Thr319fs*1 (c.955_958delACTT) (p.VL317fs) (V317fs*3), heterozygous; Mutation: RB1, p.Val654Cysfs*4 (c.1959delA), heterozygous; Mutation: SMAD4, p.Gly231Alafs*10 (c.692delG), heterozygous; Mutation: SMAD4, p.Leu495Pro (c.1484T>C), heterozygous; Mutation: TP53, p.Ser90Leufs*59 (c.267dupC) (c.267_268insC), heterozygous; Mutation: TP53, p.Tyr126Cys (c.377A>G), heterozygous

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with TrypLE Express, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO_2, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.