

## Li-7 Cells | 305102

### Informações gerais

#### Description

The Li-7 cell line is a human hepatocellular carcinoma (HCC) cell line that is commonly used in cancer research, particularly in the study of liver cancer. Derived from a primary liver tumor, Li-7 cells exhibit the typical characteristics of HCC, including the ability to produce alpha-fetoprotein (AFP), a marker often elevated in liver cancer. These cells are also known for their genetic stability, which makes them a reliable model for long-term studies.

Genomic analysis of Li-7 cells has revealed various chromosomal abnormalities that are characteristic of HCC, including gains in regions such as 5p, 8q, and 11q, and losses in 13q and 14q. These chromosomal changes are indicative of the complex genetic alterations that drive hepatocarcinogenesis. Specifically, the gain in 8q is associated with the amplification of the MYC oncogene, which plays a crucial role in cell cycle progression and proliferation, further emphasizing the utility of Li-7 cells in oncogenic pathway studies.

Li-7 cells also serve as a valuable model for studying the molecular mechanisms underlying HCC, including the pathways involving key genes like TFDP1, CUL4A, and CDC16, which have been identified as targets of amplification in HCC. These genes are involved in cell cycle regulation and DNA repair, processes that are often dysregulated in cancer. Thus, the Li-7 cell line is instrumental in elucidating the molecular events that lead to the development and progression of liver cancer, providing insights that could guide therapeutic strategies.

**Organism** Human

**Tissue** Liver

**Disease** Adult hepatocellular carcinoma

**Synonyms** LI7, Li7, C-Li-7

### Características

**Age** 45 years

**Gender** Male

**Ethnicity** Asian

**Morphology** Epithelial

**Growth properties** Adherent

### Dados regulatórios

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<b>Citation</b>	Li-7 (Cytion catalog number 305102)
<b>NCBI_TaxID</b>	9606
<b>CellosaurusAccession</b>	CVCL_3840

### Dados biomoleculares

#### Manuseio

<b>Culture Medium</b>	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO <sub>3</sub> (Cytion article number 820700a)
<b>Supplements</b>	Supplement the medium with 10% FBS
<b>Dissociation Reagent</b>	Accutase
<b>Subculturing</b>	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
<b>Freeze medium</b>	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below  $-150^{\circ}\text{C}$  to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a  $37^{\circ}\text{C}$  water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at  $300 \times g$  for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

### Incubation Atmosphere

$37^{\circ}\text{C}$ , 5%  $\text{CO}_2$ , humidified atmosphere.

### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately  $-78^{\circ}\text{C}$  throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about  $-150$  to  $-196^{\circ}\text{C}$ . Storage at  $-80^{\circ}\text{C}$  is acceptable only as a short interim step before transfer to liquid nitrogen.

## Controle de Qualidade e Análise Molecular

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**Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.