Product sheet

TPC-1 Cells | 305054

General information

Description	The TPC-1 cell line originates from a papillary thyroid carcinoma (PTC) and is widely utilized as a model for studying the molecular mechanisms of thyroid cancer. This cell line is notable for harboring the RET/PTC1 rearrangement, a hallmark genetic alteration in PTC. The RET/PTC1 fusion results in constitutive activation of RET tyrosine kinase signaling, driving oncogenic processes such as increased cellular proliferation, survival, and differentiation. This genetic feature has made TPC-1 a valuable tool in understanding thyroid oncogenesis and in evaluating targeted therapies. Derived from a well-differentiated thyroid tumor, TPC-1 retains epithelial characteristics and exhibits features associated with thyroid differentiation, including thyroglobulin production. TPC-1 has been extensively studied for its signaling pathways, particularly the MAPK and PI3K/AKT pathways, which are activated downstream of RET/PTC1. These pathways are critical to thyroid tumor progression and represent targets for therapeutic intervention. In addition to its genetic and cellular characteristics, TPC-1 has been employed in in vitro and in vivo models to investigate the effectiveness of RET inhibitors and other targeted therapies. Its well-characterized genetic background and responsiveness to pharmacological agents make it a crucial model for translational research in thyroid cancer. Studies comparing TPC-1 with other thyroid cancer cell lines have also highlighted its role in identifying common and distinct molecular features of thyroid cancer subtypes, aiding in the development of personalized treatment strategies.
Organism	Human
Tissue	Thyroid
Disease	Thyroid gland papillary carcinoma
Synonyms	TPC1

Characteristics

Age	Adult
Gender	Female
Morphology	Epithelial
Growth properties	Adherent

Regulatory Data

TPC-1 Cells | 305054

Citation	TPC-1 (Cytion catalog number 305054)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_6298

Biomolecular Data

Handling

Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with 10% FBS, 4.5 g/L Glucose
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.