

NS3-CMPK-hLBR1TM-mEGFP Cells | 300986**General information**

Description	This clonal stable cell line was generated by transfection of a circular plasmid and by Flp recombination followed by drug resistance selection.
Organism	Human
Tissue	Endocervix
Disease	Adenocarcinoma
Metastatic site	Primary tumor site (endocervix/cervix)
Applications	Nuclear envelope remodeling; lamin B receptor (LBR) biology; doxycycline-inducible gene expression; HeLa Kyoto FlpIn TReX system; chromatin/lamina interactions; live-cell imaging; conditional depletion studies using NS3-mediated proximity labeling
Synonyms	HeLa R19 FlpIn TReX H2B-Cherry/NS3-CMPK-hLBR1TM-mEGFP

Characteristics

Age	30 years
Gender	Female
Ethnicity	African American
Morphology	Epithelial-like
Cell type	Epithelial cells
Growth properties	Adherent

Regulatory Data

Citation	NS3-CMPK-hLBR1TM-mEGFP (Cytion catalog number 300986)
Biosafety level	1
NCBI_TaxID	9606

NS3-CMPK-hLBR1TM-mEGFP Cells | 300986**CellosaurusAccession** CVCL_UR51**GMO Status** GMO-S1: This HeLa R19 FlpIn TREx derivative contains Flp recombinase-integrated constructs encoding H2B-mCherry and doxycycline-inducible NS3-CMPK-hLBR1TM-mEGFP, with G418 resistance marker. This classification applies only within Germany and may differ elsewhere.**Biomolecular Data****Protein expression** H2B-mCherry and DOx inducible NS3-CMPK-hLBR1TM-mEGFP**Handling****Culture Medium** DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO₃, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)**Supplements** Supplement the medium with 10% FBS, 0.5 mg/mL G418**Dissociation Reagent** Accutase**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Fluid renewal** 2 to 3 times per week**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.