Product sheet

HK-CRISPR-Nup188-mEGFP Cells | 300657

General information

Description	HK-CRISPR-Nup188-mEGFP is a genetically modified Hela Kyoto cell line. It uses CRISPR/Cas9 technology to integrate mEGFP (monomeric Enhanced Green Fluorescent Protein) at the Nup188 locus. Nup188 is a key nucleoporin protein involved in the nuclear pore complex, essential for molecule transport between the nucleus and cytoplasm. The mEGFP tagging allows real-time visualization of Nup188, aiding in the study of nuclear pore complex dynamics and function. This cell line is particularly useful for researchers focusing on nuclear transport and nuclear pore complex biology. Its fluorescent tagging enables observation of Nup188 under various conditions, including drug treatments and genetic modifications. Applications include high-content screening, live-cell imaging, and other fluorescence-based techniques, providing valuable insights into nucleocytoplasmic transport and nuclear envelope integrity.
Organism	Human
Tissue	Endocervix
Disease	Adenocarcinoma
Metastatic site	Not applicable (HeLa Kyoto derivative; primary tumor site is endocervix)
Applications	Nuclear pore complex (NPC) biology; nucleocytoplasmic transport; Nup188 dynamics and function; live-cell fluorescence imaging of nuclear envelope; high-content screening; CRISPR knock-in validation studies; nuclear transport inhibitor evaluation

Characteristics

Age	30 years
Gender	Female
Ethnicity	African American
Morphology	Epithelial-like cells with mosaic stone shape
Cell type	Epithelial cells
Growth properties	Adherent

Regulatory Data

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Citation	HK-CRISPR-Nup188-mEGFP (Cytion catalog number 300657)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	Not assigned (HK-CRISPR-Nup188-mEGFP is a CRISPR-modified HeLa Kyoto derivative; parental HeLa Kyoto CVCL_1922)
Depositor	The Ellenberg Lab (EMBL)
GMO Status	GMO-S1: This HeLa Kyoto line contains a CRISPR knock-in of mEGFP at the Nup188 locus, enabling visualization of nuclear pore scaffold dynamics. This classification applies only within Germany and may differ elsewhere.

Biomolecular Data

Protein expression	Nup188, mEGFP-tag

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO_2, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.