Product sheet

CV-1 Cells | 605471

General information

Description	CV-1 is a African green monkey cell line derived from the kidney in 1964. Initially used in research that focused on the transformation of the cancerogenic Rous sarcoma virus (RSV), this fibroblast-like cell line is widely used in biological research for virus production, transfection, and gene silencing. These cells are negative for reverse transcriptase and being susceptible to several viruses, including poliovirus 1, herpes simplex, simian virus 40 (SV40), California encephalitis, and both Eastern and Western equine encephalitis. The CV-1 cell line exhibits rapid growth, grows adherent on plastic and glass surfaces and shows chromosome number shifts at high passage levels. It has been observed that CV-1 cells exhibit increased tumorigenicity in Wistar rats treated with ATG as well as increased cell colony formation in soft agar. Moreover, CV-1 cells support the replication of SV40 virus and exhibit rapid thymidine kinase (TK) activity following induction of simian, adeno, and papovavirus infections. The karyotype of CV-1 cells is 2n = 60, pseudodiploid. CV-1 cells have been used in a variety of specific applications in biological research, including efficacy testing, transfection host, and viruscide testing. They are also known to be a suitable host for transfection, especially by SV40 vectors.
Organism	Monkey
Tissue	Kidney
Applications	Suitable host for transfection, especially by SV40 vectors.
Synonyms	Cv-1, CV 1, CV-1.K, CV1

Characteristics

Age	141 days
Gender	Male
Cell type	Fibroblast
Growth properties	Adherent

Regulatory Data

Citation	CV-1 (Cytion catalog number 605471)

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Biosafety level	1
NCBI_TaxID	9534
CellosaurusAccession	CVCL_0229

Biomolecular Data

Virus susceptibility	Poliovirus 1, herpes simplex, Eastern equine encephalitis, Western equine encephalitis, California encephalitis, SV40
Reverse transcriptase	Negative

Handling

Culture Medium	EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO3, w: EBSS (Cytion article number 820100a)
Supplements	Supplement the medium with 10% FBS and 1% NEAA
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	3 to 4 x 10^4 cells/cm^2 will yield in a confluent layer in about 4 days
Fluid renewal	2 times per week
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.