Product sheet

U2OS-CRISPR-TPR-SNAP Cells | 300667

General information

Description	U2OS-CRISPR-TPR-SNAP is a genome-edited human osteosarcoma cell line derived from U2OS cells in which the endogenous TPR (Translocated Promoter Region) gene has been modified using CRISPR/Cas9 technology to encode an in-frame SNAP tag. TPR is a large coiled-coil nucleoporin that localizes to the nuclear basket on the nucleoplasmic side of the nuclear pore complex (NPC). By tagging TPR at its endogenous locus, the fusion protein is expressed under native regulatory control, preserving physiological expression levels and maintaining proper incorporation into the nuclear basket structure. The SNAP tag enables covalent labeling of TPR with benzylguanine-conjugated fluorescent substrates in live or fixed cells, allowing highly specific and stable visualization. In U2OS-CRISPR-TPR-SNAP cells, labeled TPR displays a characteristic punctate ring-like distribution at the nuclear envelope, corresponding to NPC-associated nuclear basket structures. This system is well suited for quantitative fluorescence microscopy, super-resolution imaging, pulse-chase labeling, and dynamic studies of nuclear basket assembly and turnover. The flat morphology and large nuclei of U2OS cells facilitate high-resolution imaging of nuclear envelope-associated structures. TPR plays critical roles in mRNA export, nuclear transport regulation, chromatin organization at the nuclear periphery, and spatial genome organization. TPR is also implicated in the formation of nuclear transport-related subcompartments and in the exclusion of heterochromatin from nuclear pore-associated regions. U2OS-CRISPR-TPR-SNAP provides a physiologically relevant model for dissecting the architecture and dynamics of the nuclear basket, investigating nucleocytoplasmic trafficking mechanisms, and studying nuclear envelope-associated chromatin interactions under endogenous expression conditions.
Organism	Human
Tissue	Bone
Disease	Osteosarcoma
Metastatic site	Primary tumor site (bone)
Applications	Nuclear basket biology; TPR-mediated mRNA export; nucleocytoplasmic transport regulation; chromatin organization at the nuclear periphery; nuclear transport subcompartments; genome spatial organization; super-resolution microscopy; SNAP pulse-chase labeling; heterochromatin exclusion from pore-associated regions

Characteristics

Age	15 years
Gender	Female
Ethnicity	Caucasian

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Morphology	Epithelial-like
Cell type	Epithelial cells (osteosarcoma)
Growth properties	Adherent

Regulatory Data

Citation	U2OS-CRISPR-TPR-SNAP (Cytion catalog number 300667)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	Not assigned (CRISPR-modified U2OS derivative; parental U2OS CVCL_0042)
Depositor	The Ellenberg Lab (EMBL)
GMO Status	GMO-S1: This human osteosarcoma cell line (U2OS-CRISPR-TPR-SNAP) contains a CRISPR-engineered TPR-SNAP fusion enabling fluorescent and chemical labeling of the TPR nuclear basket protein. The construct is stably integrated. This classification applies only within Germany and may differ elsewhere.

Biomolecular Data

Protein expression	TPR, SNAP-tag

Handling

Culture Medium	McCoys 5a, w: 3.0 g/L Glucose, w: stable Glutamine, w: 2.0 mM Sodium pyruvate, w: 2.2 g/L NaHCO3 (Cytion article number 820200a)
Supplements	Supplement the medium with 10% FBS, 3.0 g/L Glucose, stable Glutamine, 2.0 mM Sodium pyruvate, 2.2 g/L NaHCO3, 1% NEAA
Dissociation Reagent	Accutase
Doubling time	approx. 24 to 36 hours

U2OS-CRISPR-TPR-SNAP Cells | 300667

Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1 to 3
Seeding density	1 to 3 × 10^4 cells/cm^2
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

U2OS-CRISPR-TPR-SNAP Cells | 300667

Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.