Product sheet

JEG-3 Cells | 300222

General information

Description	The JEG-3 cell line is derived from a human choriocarcinoma, a type of cancer that originates from trophoblastic cells in the placenta. These cells exhibit properties characteristic of trophoblasts, including the ability to produce hormones such as human chorionic gonadotropin (hCG), which is crucial for pregnancy maintenance. JEG-3 cells are epithelial in nature and are often utilized in research focused on placental function, cancer biology, and endocrine signaling. JEG-3 cells are known for their aggressive growth characteristics and capacity to invade surrounding tissues, making them a valuable model for studying the mechanisms of trophoblastic tumor invasion and metastasis. Additionally, they have been used extensively in research investigating the molecular pathways involved in placental development, as well as the role of trophoblasts in immune tolerance during pregnancy. The cells are typically cultured in RPMI-1640 medium supplemented with fetal bovine serum and other growth factors to support their proliferation and maintenance. This cell line provides a robust platform for investigating placental cancer biology, hormone production, and the interaction between trophoblasts and the maternal immune system.
Organism	Human
Tissue	Placenta
Disease	Choriocarcinoma
Metastatic site	Brain
Applications	Transfection host
Synonyms	Jeg-3, JEG3, Jeg3, jeg3

Characteristics

Age	Fetus
Gender	Male
Morphology	Epithelial-like
Growth properties	Adherent

Regulatory Data

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Citation	JEG-3 (Cytion catalog number 300222)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_0363

Biomolecular Data

Isoenzymes	PGM3, 1-2, PGM1, 1, ES-D, 1, AK-1, 1, GLO-1, 1-2, G6PD, type B
Tumorigenic	Forms malignant tumor consistent with choriocarcinoma
Products	HCG, human chorionic somatomammotrophin (placental lactogen), progesterone.

Handling

Culture Medium	EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO3, w: EBSS (Cytion article number 820100a)
Supplements	Supplement the medium with 10% FBS and 1% NEAA
Dissociation Reagent	Accutase
Doubling time	36 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	2 x 10^4 cells/cm^2 will result in a confluent monolayer within 2 to 3 days.
Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	Allow the cells to recover from the freezing process for 24 to 48 hours.

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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Flask Coating	None
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.