**Product sheet**

# MEG-01 Cells | 300482

## General information

| | |
|---|---|
| **Description** | The MEG-01 cell line is a human megakaryoblast cell line established from the bone marrow of a 55-year-old male patient who was in the megakaryoblastic crisis phase of Chronic Myelogenous Leukemia (CML). This cell line was developed in 1983 at Nagoya University School of Medicine, Japan. The patient from whom MEG-01 was derived was positive for the Philadelphia chromosome (Ph1), a hallmark of CML. The MEG-01 cells exhibit a hyperdiploid karyotype with a modal chromosome number of 56 to 58, consistently showing the presence of the Ph1 chromosome, which is a result of the t(9;22) chromosomal translocation.<br><br>MEG-01 cells have mixed growth properties, demonstrating both adherent and suspension characteristics in culture. These cells express several markers and antigens characteristic of megakaryocytic lineage, including CD41, CD61, and CDw14. They also test positive for cytoplasmic Factor VIII, surface GPIIb/IIIa, and various enzymatic activities such as periodic acid-Schiff (PAS) reaction, alpha naphthyl acetate esterase, and acid phosphatase. Interestingly, MEG-01 cells are negative for myeloperoxidase, alpha naphthyl butyrate esterase, naphthol AS-D chloroacetate esterase, and alkaline phosphatase, which helps distinguish them from other myeloid cells.<br><br>MEG-01 has been a valuable model for studying human megakaryopoiesis, platelet production, and the biosynthesis of proteins unique to the megakaryocytic lineage, such as platelet-derived growth factor (PDGF) and glycoproteins like GPIIb/IIIa. Due to its well-characterized genetic background and its ability to express key megakaryocyte markers, MEG-01 serves as a significant tool in the investigation of leukemia and platelet biogenesis mechanisms, although it is not intended for therapeutic or in vivo applications. |
| **Organism** | Human |
| **Tissue** | Bone marrow |
| **Disease** | Chronic myeloid leukemia |
| **Synonyms** | Meg-01, MEG01, Meg01 |

## Characteristics

| | |
|---|---|
| **Age** | 55 years |
| **Gender** | Male |
| **Ethnicity** | East Asian |
| **Morphology** | Myoblast-like |
| **Cell type** | Megakaryoblast |

# MEG-01 Cells | 300482

| | |
|---|---|
| **Growth properties** | Adherent/suspension |

## Regulatory Data

| | |
|---|---|
| **Citation** | MEG-01 (Cytion catalog number 300482) |
| **Biosafety level** | 1 |
| **NCBI_TaxID** | 9606 |
| **CellosaurusAccession** | CVCL_0425 |

## Biomolecular Data

| | |
|---|---|
| **Antigen expression** | CD41 +, CD61 +, CDw14 + |

## Handling

| | |
|---|---|
| **Culture Medium** | RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a) |
| **Supplements** | Supplement the medium with 10% FBS |
| **Dissociation Reagent** | Accutase |
| **Subculturing** | Gather the suspension cells in a 15 ml tube and gently wash the adherent cells with PBS lacking calcium and magnesium (use 3-5 ml for T25 flasks and 5-10 ml for T75 flasks). Apply Accutase (1-2 ml for T25 flasks, 2.5 ml for T75 flasks) ensuring full coverage of the cell layer. Allow the cells to incubate at room temperature for 10 minutes. Following incubation, combine and centrifuge both the suspension and adherent cells. After centrifugation, carefully resuspend the cell pellet and transfer the cell suspension into new flasks containing fresh medium. |
| **Freeze medium** | As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress. |

| | |
|---|---|
| **Thawing and Culturing Cells** | 1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.<br>2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.<br>3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.<br>4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.<br>5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.<br>6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.<br>7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.<br>8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes. |
| **Incubation Atmosphere** | 37°C, 5% $CO_2$, humidified atmosphere. |
| **Shipping Conditions** | Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage. |
| **Storage Conditions** | For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen. |

## Quality Control & Molecular Analysis

| | |
|---|---|
| **Sterility** | Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.<br><br>To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections. |