

V79-GalK1 Cells | 305011**General information**

Description	This cell line is a clonal derivative of V79 cells that had been treated with ethylmethanesulfonate. The cells are deficient in galactokinase and are resistant to 2-deoxygalactose and 8-azaguanine.
Organism	Chinese hamster
Tissue	Lung
Disease	Normal lung fibroblast (galactokinase-deficient; EMS-mutagenized V79 derivative; resistant to 2-deoxygalactose and 8-azaguanine)
Metastatic site	Not applicable (Chinese hamster lung fibroblast cell line; not a tumor sample)
Applications	Mutation research; genetic toxicology; genotoxicity testing; galactokinase deficiency complementation studies; HPRT-deficient parent V79 genetics; CHO/V79 model comparisons; DNA repair research

Characteristics

Gender	Male
Ethnicity	Not applicable (Chinese hamster cell line)
Morphology	Fibroblast-Like
Cell type	Fibroblast
Growth properties	Adherent

Regulatory Data

Citation	V79-GalK1 (Cytion catalog number 305011)
Biosafety level	1
NCBI_TaxID	10029
CellosaurusAccession	CVCL_2797

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GMO Status	No genetic modification (mutagenesis by EMS is a chemical mutagen treatment, not stable transgene introduction); carries galactokinase deficiency and 8-azaguanine resistance arising from endogenous mutations
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Biomolecular Data

Karyotype	2n=22
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Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO ₃ , w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
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Supplements	Supplement the medium with 10% FBS
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Dissociation Reagent	Accutase
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Doubling time	approx. 14 to 18 hours (V79-derived; rapid doubling)
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Split ratio	1 to 5
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Seeding density	2-4*10 ⁴ /cm ²
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Fluid renewal	2 to 3 times per week
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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
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V79-GalK1 Cells | 305011**Thawing and
Culturing Cells**

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

**Incubation
Atmosphere**

37°C, 5% CO₂, humidified atmosphere.

**Shipping
Conditions**

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

**Storage
Conditions**

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.