Product sheet

MALME-3M Cells | 305583

General information

Description	The MALME-3M cell line is a human melanoma model used extensively in cancer research to investigate mechanisms of melanoma progression, immune evasion, and drug resistance. This cell line is derived from a metastatic melanoma lesion and demonstrates several characteristics relevant to aggressive melanoma, including its ability to express key oncogenic markers such as HER2 and its role in modulating the tumor microenvironment. Studies involving MALME-3M have highlighted its responsiveness to targeted therapies, such as bispecific antibodies aimed at HER2, and its use in evaluating T-cell mediated immunotherapies. One significant area of research involving MALME-3M cells is their utility in studying the mechanisms of immune evasion in melanoma. For instance, co-culture systems pairing MALME-3M with immune cells allow researchers to explore how melanoma cells modulate immune responses through pathways like PD-1/PD-L1 and other immune checkpoint inhibitors. This cell line has also been genetically modified to study the effects of gene perturbations on immune interactions, making it a valuable tool for high-throughput genetic screening. In addition to its role in immunological studies, MALME-3M cells are instrumental in exploring the effects of growth hormone (GH) on melanoma progression. Research has demonstrated that GH can enhance drug resistance and metastatic potential in MALME-3M cells by altering the composition of melanoma-derived exosomes. These exosomes can transfer drug resistance and migration-promoting factors to other cells in the tumor microenvironment. Such studies underscore the potential of targeting GH signaling pathways as a therapeutic strategy for overcoming melanoma chemoresistance.
Organism	Human
Tissue	Skin
Disease	Melanoma
Metastatic site	Lung
Synonyms	Malme-3M, MALME 3M, Malme-3 M, MALME.3M, Malme3M, MALME3M, Malme-3 Monolayer

Characteristics

Age	43 years
Gender	Male
Ethnicity	Caucasian
Morphology	Fibroblast-like
Cell type	Fibroblast

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Growth properties	Adherent

Regulatory Data

Citation	MALME-3M (Cytion catalog number 305583)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1438

Biomolecular Data

Antigen expression	HLA A2, Aw30, B13, B40(+/-), DRw7
Tumorigenic	Yes, in nude mice

Handling

Culture Medium	IMDM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 25 mM HEPES, w: 1.0 mM Sodium pyruvate, w: 3.024 g/L NaHCO3 (Cytion article number 820800a)
Supplements	Supplement the medium with 20% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with TrypLE Express, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	3×10^4 cells/cm^2
Fluid renewal	2 to 3 times per week

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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.