Product sheet

HEK293-CXCR7 Cells | 305421

General information

Description	**Disclaimer: The prices displayed for cell lines are exclusively for academic/not-for-profit customers. For commercial entitites the price is approximately €6,250.** **If you represent a commercial entity or are unsure which category applies, please contact us.** The HEK293-CXCR7 cell line is a stable recombinant HEK293 cell line engineered to express the CXCR7 receptor at a low level. This cell line was developed using inscreenex's landing pad technology, ensuring precise and reproducible integration of the CXCR7 gene at a specific, pre-validated genomic locus. CXCR7, also known as ACKR3, is an atypical chemokine receptor that modulates immune responses and tumor biology. Unlike classical GPCRs, CXCR7 does not signal through G proteins; instead, it scavenges chemokines like CXCL12 and CXCL11 and forms heterodimers with CXCR4, contributing to tumor growth, metastasis, and poor prognosis in various cancers, including breast, lung, and prostate cancers. The expression of CXCR7 in this cell line was confirmed using flow cytometry with a target-specific antibody, ensuring reliable expression across the cell population. However, the receptor density was not quantified in this cell line.
Organism	Human
Tissue	Fetal Kidney
Disease	Transformed/immortalized; non-tumorigenic (HEK293 background)
Applications	CXCR7/ACKR3-targeted antibody development; GPCR pharmacology; chemokine receptor biology; CXCL11/CXCL12 signaling studies; oncology and cardiovascular research

Characteristics

Age	Fetus
Gender	Female
Morphology	Epithelial-like
Cell type	Epithelial cells
Growth properties	Monolayer, adherent

Regulatory Data

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Citation	HEK293-CXCR7 (Cytion catalog number 305421)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_6G22
GMO Status	GMO-S1: This HEK293 line contains a CXCR7 expression construct, enabling studies of chemokine receptor activity. This classification applies only within Germany and may differ elsewhere.

Biomolecular Data

Receptors expressed	CXCR7 (ACKR3)

Handling

Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with 10% FBS, 1 mM sodium pyruvate, 10 mM HEPES, 1% NEAA. Add Geneticin (G418-Sulfat) to achieve a final concentration of 1 mg/mL.
Dissociation Reagent	Trypsin-EDTA
Doubling time	approx. 24-36 hours
Subculturing	For routine adherent cell culture: Aspirate the old culture medium from the adherent cells, and wash them with PBS to remove any remaining medium. After aspirating the PBS, add the appropriate volume of Trypsin/EDTA solution based on the culture vessel size (e.g., 1 ml for a T25 flask, 3 ml for a T75 flask) and incubate at room temperature or 37°C until the cells detach (5-10 minutes). Monitor detachment under a microscope, and gently tap the vessel if necessary to release the cells. Once detached, add complete medium to inactivate the Trypsin/EDTA, gently resuspend the cells, and transfer an aliquot of the cell suspension into a new culture vessel containing fresh medium. Place the vessel in an incubator set to 37°C with 5% CO_2, and change the medium every 2-3 days.
Split ratio	1 to 5
Seeding density	2 to 4 x 10^4 cells/cm^2

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Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, split the cells at a ratio of 1:2 to 1:3 in T25 flasks and allow the cells to recover from the freezing process and to adhere for at least 24 hours. For best attachment and viability after thawing the cells, we recommend using Collagen-coated flasks or plates for the initial seeding after cryo-recovery. Collagen coating is not required for subsequent routine culture of the cells.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.

Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.