

B-LCL-HROC60 | 302004

Cell line

Description	B-LCL-HROC60 is a B-cell lymphoma cell line derived from a 71-year-old female patient with B-cell lymphoma. The cell line was established from a biopsy specimen and is maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin. The cell line is characterized by its high proliferation rate and its ability to produce monoclonal antibodies. It is a B-cell lymphoma cell line derived from a patient with B-cell lymphoma. The cell line was established from a biopsy specimen and is maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin. The cell line is characterized by its high proliferation rate and its ability to produce monoclonal antibodies. It is a B-cell lymphoma cell line derived from a patient with B-cell lymphoma. The cell line was established from a biopsy specimen and is maintained in RPMI 1640 medium supplemented with 10% fetal bovine serum (FBS) and 1% penicillin-streptomycin. The cell line is characterized by its high proliferation rate and its ability to produce monoclonal antibodies.
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Organism	Human
Tissue	B lymphocyte
Disease	B-cell lymphoma
Synonyms	Bc HROC60, TiBcHROC60

Cell line characteristics

Age	71 years
Gender	Female
Ethnicity	German
Morphology	Clonal
Cell type	B-cell
Growth properties	Adherent

Cell line identification

Citation	B-LCL-HROC60 (Cytion 302004)
Biosafety level	2
NCBI_TaxID	9606
CellosaurusAccession	CVCL_A7UT

EBV-LCL-HROC60 | 302004

EBV-LCL-HROC60

Surface antigens	CD19
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Viruses	EBV
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EBV

Culture Medium	RPMI 1640, w: 2.0 mM NaH_2PO_4 , w: 2.0 g/L NaHCO_3 (Cytion 820700a)
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Supplements	10% FBS
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Subculturing	1:5
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Freeze medium	
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Thawing and Culturing Cells	1. Thaw cells quickly in a water bath at 37°C. Transfer cells to a centrifuge tube and centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cell pellet in 1 mL of culture medium. 2. Seed cells into a T25 flask containing 10 mL of culture medium. 3. Incubate cells in a humidified CO₂ incubator at 37°C. 4. Monitor cell growth and confluency. Once cells reach 70-80% confluency, passage the cells. 5. Detach cells using trypsin-EDTA. Add 1 mL of trypsin to the flask and incubate for 2-3 minutes. 6. Add 4 mL of cold medium to stop the trypsin reaction. Pipette the cells into a centrifuge tube and centrifuge at 300 x g for 5 minutes. 7. Remove the supernatant and resuspend the cell pellet in 1 mL of culture medium. 8. Seed cells into a new T25 flask containing 10 mL of culture medium.
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Incubation Atmosphere	37°C, 5% CO ₂
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Flask Coating	
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