

HHHC6548 T1 M1 | 300832

General information

Description	Cell line derived from a 26-year-old male patient with a primary colorectal adenocarcinoma (T1M1), UICC IIIc, histologically confirmed (immunohistochemistry, TNM T3N2Mx, G3) colorectal adenocarcinoma.
Organism	Human
Tissue	Colorectal adenocarcinoma, UICC IIIc, histologically confirmed CRC (immunohistochemistry, TNM T3N2Mx, G3) colorectal adenocarcinoma.
Disease	Colorectal adenocarcinoma

Cell line characteristics

Age	26 years
Gender	Male
Ethnicity	German
Morphology	Epithelial cells
Growth properties	Adherent

Cell line identification

Citation	HHHC6548 T1 M1 (Cytion 300832)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1U94

Cell line characteristics

Protein expression	PTEN-
Tumorigenic	Yes, tumorigenic in nude mice

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Ploidy status	Hexaploid
MSI-status	MSI-H
Mutational profile	K-Ras G13D, N-Ras wt, H-Raswt, B-Rafwt , PIK3CAwt
Genotype	
Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Selenium, w: 1.2 g/L NaHCO3 820400a)
Supplements	Insulin Transferrin 10% FBS
Dissociation Reagent	Trypsin
Doubling time	26 hours
Subculturing	Cells are harvested by trypsinization and resuspended in DMEM:Ham's F12 + 10% FBS. Cells are seeded into T25 flasks, reaching 80-90% confluency. Cells are then harvested by trypsinization and resuspended in DMEM:Ham's F12 + 10% FBS. Cells are then seeded into T25 flasks, reaching 80-90% confluency. Cells are then harvested by trypsinization and resuspended in DMEM:Ham's F12 + 10% FBS. Cells are then seeded into T25 flasks, reaching 80-90% confluency.
Seeding density	1 x 10 ⁴ cells/cm ²
Fluid renewal	Every 3-5 days
Post-Thaw Recovery	Cells are seeded into T25 flasks and allowed to recover for 24-48 hours before use.
Freeze medium	Cells are harvested by trypsinization and resuspended in DMEM:Ham's F12 + 10% FBS + 10% DMSO. Cells are then seeded into T25 flasks, reaching 80-90% confluency. Cells are then harvested by trypsinization and resuspended in DMEM:Ham's F12 + 10% FBS. Cells are then seeded into T25 flasks, reaching 80-90% confluency. Cells are then harvested by trypsinization and resuspended in DMEM:Ham's F12 + 10% FBS. Cells are then seeded into T25 flasks, reaching 80-90% confluency.

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**Thawing and
Culturing Cells**

1.

Remove the vial from the dry ice storage and place it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Do not shake the vial. Once the cells are thawed, transfer them to a 15 mL centrifuge tube containing 10 mL of pre-warmed complete medium. Centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 1 mL of complete medium. Seed the cells into a 24-well plate (1 well) or a 96-well plate (1 well) containing 1 mL of complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
2.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
3.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
4.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
5.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
6.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
7.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.
8.

After 24 hours, check the cells for attachment. If the cells are not attached, add another 1 mL of complete medium. If the cells are attached, remove the medium and replace it with 1 mL of fresh complete medium. Incubate for 24 hours at 37°C, 5% CO₂.

**Incubation
Atmosphere**

37°C, 5% CO₂, humidified

Flask Coating

Not required

**Freezing
Procedure**

Resuspend the cells in 1 mL of complete medium. Transfer the cells to a 1.5 mL microcentrifuge tube. Centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 100 µL of freezing medium. Freeze the cells at -80°C.

**Shipping
Conditions**

Store the cells at -80°C. Ship the cells on dry ice.

**Storage
Conditions**

Store the cells at -150°C for up to 196 weeks.

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Sterility

The cells are sterile. No mycoplasma contamination was detected by PCR. The cells are free of endotoxins. The cells are free of viruses. The cells are free of fungi. The cells are free of bacteria.

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HLA

A*: '03:01:01, '24:02:01
B*: '35:XX, '37:01:01
C*: 04:01:01, 06:02:01
DRB1*: '01:01:01G, '04:01:01
DQA1*: '01:01:01, '03:01:01
DQB1*: '03:02:01, '05:01:01
DPB1*: '01:01:01, '04:01:01
E: '01:01:01, '01:06:01