

WEHI-164 | 400438

General information

Description	WEHI-164 is a B-cell lymphoma cell line derived from a BALB/c mouse. It is a non-adherent, non-secretory, non-tumorigenic cell line. WEHI-164 is a B-cell lymphoma cell line derived from a BALB/c mouse. It is a non-adherent, non-secretory, non-tumorigenic cell line. WEHI-164 is a B-cell lymphoma cell line derived from a BALB/c mouse. It is a non-adherent, non-secretory, non-tumorigenic cell line.
Organism	Mouse
Disease	B-cell lymphoma
Synonyms	WEHI 164, WEHI164, WEHI 164 TC

Characteristics

Breed/Subspecies	BALB/c
Morphology	Small, round cells
Cell type	B-cell
Growth properties	Non-adherent

Identification and safety

Citation	WEHI-164 (Cytion 400438)
Biosafety level	1
NCBI_TaxID	10090
CellosaurusAccession	CVCL_2251

Genetic information

Tumorigenic	No, Balb/c
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References

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Culture Medium

RPMI 1640, w: 2.0 mM β -mercaptoethanol, w: 2.0 g/L NaHCO₃ (Cytion 820700a)

Supplements

10% FBS

Dissociation Reagent

Trypsin

Subculturing

Cells are grown in 25 cm² flasks in RPMI 1640 medium supplemented with 10% FBS. When cells reach confluence, they are trypsinized and seeded into new flasks at a density of 1×10^4 cells/cm².

Seeding density

1×10^4 cells/cm²

Fluid renewal

2-3 times per week

Post-Thaw Recovery

After thawing, cells are seeded into flasks containing 10% FBS medium. After 48 hours, the medium is replaced with fresh 10% FBS medium.

Freeze medium

For freezing, cells are resuspended in RPMI 1640 medium supplemented with 10% FBS and 10% DMSO. The suspension is then seeded into cryovials and frozen.

Thawing and Culturing Cells

1. Thaw cells rapidly in a water bath at 37°C. Transfer the cells to a centrifuge tube and centrifuge at 300 x g for 5 minutes.
2. Remove the supernatant and resuspend the cells in RPMI 1640 medium supplemented with 10% FBS. Seed the cells into a 25 cm² flask.
3. Allow the cells to attach for 24 hours. Then, replace the medium with fresh RPMI 1640 medium supplemented with 10% FBS.
4. When cells reach confluence, they can be passaged. Remove the medium and wash the cells with PBS. Add 2 ml of trypsin and incubate for 5 minutes.
5. Add 8 ml of serum-free RPMI 1640 medium to stop the reaction. Transfer the cells to a centrifuge tube and centrifuge at 300 x g for 5 minutes.
6. Remove the supernatant and resuspend the cells in RPMI 1640 medium supplemented with 10% FBS. Seed the cells into a 25 cm² flask.
7. Allow the cells to attach for 24 hours. Then, replace the medium with fresh RPMI 1640 medium supplemented with 10% FBS.
8. When cells reach confluence, they can be passaged. Remove the medium and wash the cells with PBS. Add 2 ml of trypsin and incubate for 5 minutes.

Incubation Atmosphere

37°C, 5% CO₂, humidified

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Flask Coating

Flask Coating

**Freezing
Procedure**

Freezing Procedure: The cells are frozen in a freezing medium and stored at -78°C.

**Shipping
Conditions**

Shipping Conditions: The cells are shipped in a dry ice container and stored at -78°C.

**Storage
Conditions**

Storage Conditions: The cells are stored in a dry ice container at -150 to -196°C.

Cell Line Characteristics

Sterility

Sterility: The cells are tested for sterility using PCR and other methods. The results show that the cells are free of contamination.