

XXXXX L-540 | 300201

XXXXX XXXXX

Description	L-540 XXXX XX XXXXX XX XXXXXXXX XXXXX XXXXX' XXX XXXX XXX, XXXXXXXX XXXXXXX XXXXXXX XXXXX XX XX XXXXX. XX XXXXX XX XXXXX XXXXXXX XXXXX XX XXXXXXX XXXXXXX XXX L-540 XXX XXXXXXX XXX CD30, XX XXXXXXX XXXXXXX XXXXX XXX XXXXXXX, XXX XXXXXXX XXXXXXX XXXXXXX XXXX XX XXXXXXX
Organism	XXXX
Tissue	XX XXXX
Disease	XXXXXXXX XXXX XXXXX XXX
Synonyms	L 540, L540

XXXXXXXXXXXX

Age	20 XXXX
Gender	XXXXX
Ethnicity	XXXXXXXXXX
Morphology	XXXXX XXXXXXX
Growth properties	XXXXXX

XXXXXXXXX XXXXXXXXXXXXXXX

Citation	L-540 (XXXXX XXXXXXX Cytion 300201)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1362

XXXXXXXXX XXXX- XXXXXXXXXXXXXXX

EBV-L-540 | 300201

Viruses

EBV

EBV-L-540

Culture Medium

RPMI 1640, w: 2.0 mM NaH_2PO_4 , w: 2.0 g/L NaHCO_3 (Cytion 820700a)

Supplements

10% FBS

Subculturing

5

Fluid renewal

3

Freeze medium

10% FBS + 10% DMSO

Thawing and Culturing Cells

1. Thaw cells rapidly in a water bath at 37°C. Transfer cells to a sterile tube and centrifuge at 300 x g for 3 minutes. Remove supernatant and resuspend cells in 10 ml of culture medium.
2. Seed cells into a T25 flask containing 10 ml of culture medium. Incubate at 37°C in 5% CO_2 .
3. Monitor cell growth and confluency. Once cells reach 70-80% confluency, passage the cells.
4. Remove medium and wash cells with PBS. Add 1 ml of trypsin-EDTA solution and incubate at 37°C for 5 minutes.
5. Add 4 ml of serum-free medium to stop the reaction. Pipette up and down to dissociate cells. Centrifuge at 300 x g for 3 minutes.
6. Resuspend cell pellet in 10 ml of culture medium. Count cells and seed into a new T25 flask.
7. Repeat the process for subsequent passages.
8. Maintain cells in culture medium with 10% FBS.

Incubation Atmosphere

37°C, 5% CO_2

Flask Coating

EBV

XXXX L-540 | 300201

Freezing Procedure

-78°C

Shipping Conditions

[illegible]

Storage Conditions

[illegible]

Sterility

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