

CHO-CXCR7 | 305412MH

General information

Description	CHO-CXCR7-Medium-high is a CHO (Chinese Hamster Ovary) cell line expressing CXCR7 (ACKR3) on its surface. The cell line is derived from CHO cells and is characterized by its high expression level of CXCR7. It is used for studying the function of CXCR7 and for developing CXCR7-targeted therapies. CHO-CXCR7-Medium-high cells are derived from CHO (Chinese Hamster Ovary) cells. They are characterized by their high expression level of CXCR7 (ACKR3) on the cell surface. The cells are used for studying the function of CXCR7 and for developing CXCR7-targeted therapies. CXCR7 is a GPCR (G-protein-coupled receptor) that is expressed on the surface of various cells, including endothelial cells, fibroblasts, and epithelial cells. It is involved in various biological processes, including chemotaxis, cell proliferation, and differentiation.
Organism	CHO
Tissue	CHO
Disease	Chinese hamster ovary, non-neoplastic; genetically engineered for CXCR7 (ACKR3) surface expression (medium-high expression level)
Applications	Antibody screening; CXCR7-targeted therapy development; chemokine receptor biology; tumor microenvironment research; flow cytometry
Synonyms	CHO-CXCR7

Cell characteristics

Age	CHO
Gender	CHO
Morphology	CHO CHO
Cell type	Epithelial cells
Growth properties	CHO/CHO

Cell culture conditions

Citation	CHO-CXCR7 (ACKR3) (CHO CHO Cytion 305412MH)
Biosafety level	1

Product sheet

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NCBI_TaxID	10029
CellosaurusAccession	CVCL_A8W2
GMO Status	GMO-S1: This CHO cell line contains a construct supporting medium-to-high expression of human CXCR7 for chemokine receptor research. This classification applies only within Germany and may differ elsewhere.
Receptors expressed	
CXCR7 (ACKR3)	
Culture Medium	
DMEM:Ham's F12 (1:1), w: 3.1 g/L L-glutamine, w: 2.5 mM L-ascorbic acid, w: 15 mM HEPES, w: 0.5 mM beta-mercaptoethanol, w: 820400a) CHO Growth Medium A (InSCREENeX; InSCREENeX INS-ME-1039)	
Supplements	5% FBS. Geneticin (G418-Sulfat) 0.5 mg/ml
Dissociation Reagent	EDTA
Doubling time	approx. 14-16 hours
Subculturing	Cells are detached using trypsin and seeded into new flasks. Media is replaced with fresh medium. Cells are grown in CO2, 2-3 passages.
Split ratio	1 to 5
Seeding density	2 to 5 x 10 ⁴ cells/cm ²
Fluid renewal	2-3 times per week
Post-Thaw Recovery	Cells are thawed and seeded into flasks. Media is replaced with fresh medium. Cells are grown in CO2, 2-3 passages.
Freeze medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L L-glutamine, w: 2.5 mM L-ascorbic acid, w: 15 mM HEPES, w: 0.5 mM beta-mercaptoethanol, w: 820400a) CHO Growth Medium A (InSCREENeX; InSCREENeX INS-ME-1039) + 10% DMSO

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**Thawing and
Culturing Cells**

1. Thaw cells rapidly in a water bath at 37 °C. Transfer the cells to a sterile tube and centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 ml of complete medium.
2. Seed cells into a 24-well plate at a density of 150,000 cells per well. Incubate at 37 °C in 5% CO₂ for 24 hours.
3. After 24 hours, replace the medium with fresh complete medium.
4. After 48 hours, replace the medium with fresh complete medium.
5. After 72 hours, replace the medium with fresh complete medium.
6. After 96 hours, replace the medium with fresh complete medium.
7. After 120 hours, replace the medium with fresh complete medium.
8. After 144 hours, replace the medium with fresh complete medium.

**Incubation
Atmosphere**

37 °C, 5% CO₂, humidified atmosphere.

**Shipping
Conditions**

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

**Storage
Conditions**

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

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Sterility

Cells are tested for mycoplasma contamination using PCR. Cells are also tested for endotoxin contamination using a Limulus amoebocyte lysate (LAL) assay.