

CHO-CXCR7 | 305412L

General information

Description	CHO-CXCR7-Medium-high is a CHO (Chinese Hamster Ovary) cell line expressing CXCR7 (ACKR3) on its surface. The cells are adapted for growth in DMEM/F12 medium supplemented with 10% FBS and 10% HEPES. CXCR7 is a GPCR that binds to CXCL12 and CXCL16, and is involved in chemotaxis and cell migration. The cells are suitable for antibody screening, CXCR7-targeted therapy development, chemokine receptor biology, tumor microenvironment research, and flow cytometry.
Organism	CHO
Tissue	CHO
Disease	Chinese hamster ovary, non-neoplastic; genetically engineered for CXCR7 (ACKR3) surface expression (low expression level)
Applications	Antibody screening; CXCR7-targeted therapy development; chemokine receptor biology; tumor microenvironment research; flow cytometry
Synonyms	CHO-CXCR7

Cell characteristics

Age	CHO
Gender	CHO
Morphology	CHO CHO
Cell type	Epithelial cells
Growth properties	CHO/CHO

Identification and authentication

Citation	CHO-CXCR7 CHO-CHO (CHO CHO Cytion 305412MH)
Biosafety level	1

NCBI_TaxID	10029
CellosaurusAccession	CVCL_A8W1
GMO Status	GMO-S1: This CHO cell line contains a recombinant CXCR7 expression cassette at low levels, suitable for controlled receptor-ligand studies. This classification applies only within Germany and may differ elsewhere.

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Receptors expressed	CXCR7 (ACKR3)
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Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L L-Ascorbic acid, w: 2.5 mM L-Ascorbic acid, w: 15 mM HEPES, w: 0.5 mM β-mercaptoethanol, w: 820400a) CHO Growth Medium A (InSCREENeX; InSCREENeX INS-ME-1039)
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Supplements □□□□□□□□ □□□□□ □□□□ □□□□□ 5% FBS. □□□□□ Geneticin (G418-Sulfat) □□□ □□□□□ □□□□□ □□□□ □□ 0.5 μl/μl/μl.

Dissociation Reagent $\text{Na}_2\text{S}_2\text{O}_8$ $\text{Na}_2\text{S}_2\text{O}_5$ $\text{Na}_2\text{S}_2\text{O}_4$ -EDTA

Doubling time approx. 14-16 hours

[illegible]

Split ratio 1 to 5

Seeding density	2 to 5 x 10 ⁴ cells/cm ²
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Fluid renewal 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56 57 58 59 60 61 62 63 64 65 66 67 68 69 70 71 72 73 74 75 76 77 78 79 80 81 82 83 84 85 86 87 88 89 90 91 92 93 94 95 96 97 98 99 100

Post-Thaw Recovery (10 min)

1. **Thawing:** Thaw the sample in a water bath at 37°C for 10 min. **DO NOT** vortex or shake the sample.

2. **Centrifugation:** Centrifuge the sample at 14,000 rpm for 1 min at 4°C. **DO NOT** vortex or shake the sample.

3. **Transfer:** Carefully transfer the supernatant to a clean vial. **DO NOT** vortex or shake the sample.

4. **Storage:** Store the sample at -80°C for long-term storage. **DO NOT** vortex or shake the sample.

Freeze medium α -MEM (Gibco), 10% FBS (Gibco) + 10% DMSO (Gibco) α -MEM (Gibco), 10% FBS (Gibco) + 10% DMSO (Gibco), CM-1 (Gibco)

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**Thawing and
Culturing Cells**

1. Thaw the vial rapidly in a water bath at 37°C. Remove the vial and transfer the cells to a centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 ml of complete medium.
2. Seed the cells into a 24-well plate at a density of 150,000 cells per well. Incubate at 37°C in a humidified atmosphere with 5% CO₂.
3. After 24 hours, replace the medium with fresh complete medium.
4. After 48 hours, replace the medium with fresh complete medium.
5. After 72 hours, replace the medium with fresh complete medium.
6. After 96 hours, replace the medium with fresh complete medium.
7. After 120 hours, replace the medium with fresh complete medium.
8. After 144 hours, replace the medium with fresh complete medium.

**Incubation
Atmosphere**

37°C, 5% CO₂, humidified atmosphere.

**Shipping
Conditions**

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

**Storage
Conditions**

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

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Sterility

The cells are tested for mycoplasma contamination using PCR. The results are provided in the Certificate of Analysis (COA).