

Product sheet

OVCA5 | 305616

Mutational profile KRAS, p.Gly12Val (c.35G>T),

Culture Medium RPMI 1640, w: 2.0 mM, w: 2.0 g/L NaHCO3 (Cytion 820700a)

Supplements 10% FBS

Dissociation Reagent

Doubling time 27

Split ratio 1:5

Fluid renewal 2-3

Freeze medium (FBS) + 10% DMSO

Thawing and Culturing Cells

1. Thaw cells rapidly in a 37°C water bath, then transfer to a 15 mL centrifuge tube containing 10 mL of pre-warmed complete medium. Gently resuspend cells and centrifuge at 300 x g for 5 minutes. Remove supernatant and resuspend cells in 1 mL of complete medium. Seed cells into a 24-well plate (100,000 cells/well) or a 96-well plate (20,000 cells/well).
2. Incubate cells in a humidified 5% CO2 incubator at 37°C. Monitor cell growth and confluency.
3. Once cells reach 70-80% confluency, passage them into fresh medium.
4. For passage, trypsinize cells and resuspend in 1 mL of complete medium. Seed into a 24-well plate (100,000 cells/well) or a 96-well plate (20,000 cells/well).
5. Repeat steps 1-4 as needed.
6. For freezing, trypsinize cells and resuspend in 1 mL of freezing medium. Seed into a 24-well plate (100,000 cells/well) or a 96-well plate (20,000 cells/well).
7. Incubate cells in a humidified 5% CO2 incubator at 37°C. Monitor cell growth and confluency.
8. Once cells reach 70-80% confluency, passage them into fresh medium.

