

XXX HCC1395 | 305546

Description	肝細胞性肝癌 HCC1395 細胞株は、肝臓がんのモデルとして広く用いられる。この細胞株は、1995年に、肝臓がん患者の手術で得られた組織から分離された。HCC1395 細胞株は、肝臓がんの発生、進展、転移のメカニズムを研究するための有用なツールである。この細胞株は、肝臓がんの発生、進展、転移のメカニズムを研究するための有用なツールである。この細胞株は、肝臓がんの発生、進展、転移のメカニズムを研究するための有用なツールである。
Organism	ヒト
Tissue	肝臓
Disease	肝臓がん
Synonyms	HCC-1395, SCC-1395, 肝細胞性肝癌細胞株 1395

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Age	43 男男男
Gender	男男男
Ethnicity	男男男男男
Morphology	男男男 男男男男
Cell type	男 男男男男
Growth properties	男男

Citation	HCC1395 (细胞系 细胞系 Cytion 305546)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1249

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**Thawing and
Culturing Cells**

1.

Remove the vial from liquid nitrogen storage and allow it to warm to room temperature. Do not shake the vial. Transfer the cells to a 15 mL conical centrifuge tube and centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of complete growth medium. Seed the cells into a 25 cm² flask containing 10 mL of complete growth medium.
2.

Incubate the cells at 37°C in 5% CO₂. The cells should reach confluence within 7-10 days. Once confluent, the cells can be passaged.
3.

For passage, remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of complete growth medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of complete growth medium. Seed the cells into a 25 cm² flask containing 10 mL of complete growth medium.
4.

For freezing, remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of freezing medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of freezing medium. Seed the cells into a 1.5 mL microcentrifuge tube and freeze at -80°C.
5.

For long-term storage, remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of freezing medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of freezing medium. Seed the cells into a 1.5 mL microcentrifuge tube and freeze at -80°C.
6.

For thawing, transfer the vial to a 37°C water bath and allow it to warm. Add 8 mL of complete growth medium to stop the reaction. Transfer the cells to a 15 mL conical centrifuge tube and centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of complete growth medium. Seed the cells into a 25 cm² flask containing 10 mL of complete growth medium.
7.

For passaging, remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of complete growth medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of complete growth medium. Seed the cells into a 25 cm² flask containing 10 mL of complete growth medium.
8.

For freezing, remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of freezing medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of freezing medium. Seed the cells into a 1.5 mL microcentrifuge tube and freeze at -80°C.

**Incubation
Atmosphere**

37°C, 5% CO₂, humidified

Flask Coating

Not required

**Freezing
Procedure**

Remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of freezing medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of freezing medium. Seed the cells into a 1.5 mL microcentrifuge tube and freeze at -80°C.

**Shipping
Conditions**

Remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of freezing medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of freezing medium. Seed the cells into a 1.5 mL microcentrifuge tube and freeze at -80°C.

**Storage
Conditions**

Remove the medium and wash the cells with PBS. Add 2 mL of trypsin-EDTA solution and incubate for 5 minutes. Add 8 mL of freezing medium to stop the reaction. Detach the cells by gentle pipetting and transfer to a 15 mL conical centrifuge tube. Centrifuge at 300 x g for 5 minutes. Remove the supernatant and resuspend the cells in 1 mL of freezing medium. Seed the cells into a 1.5 mL microcentrifuge tube and freeze at -80°C.

Additional Information

Sterility

Cells are provided as a suspension in complete growth medium. The cells are not tested for mycoplasma contamination. The cells are not tested for endotoxin contamination. The cells are not tested for viral contamination. The cells are not tested for bacterial contamination. The cells are not tested for fungal contamination. The cells are not tested for parasitic contamination. The cells are not tested for any other contamination.