

DT40 | 305249

General information

Description	DT40 is a B cell line derived from a chicken (Gallus gallus) embryo. It is a continuous cell line that can be grown in the presence of B cell growth factors. DT40 is a B cell line that is used for the study of B cell development and differentiation. DT40 is a B cell line that is used for the study of B cell development and differentiation.
Organism	Chicken
Disease	DT40 is a B cell line that is used for the study of B cell development and differentiation.
Synonyms	DT-40, DT 40, DT40B, LSCC-DT40

Cell characteristics

Breed/Subspecies	Hy-Line SC
Age	1 day old
Gender	Female
Morphology	Small, round cells
Cell type	B
Growth properties	DT40 is a B cell line that is used for the study of B cell development and differentiation.

Identification and safety

Citation	DT40 (Cytion 305249)
Biosafety level	1
NCBI_TaxID	9031
CellosaurusAccession	CVCL_0249

Additional information

Product sheet

DT40 | 305249

Receptors expressed	CD133, CD133 ⁺ M
Protein expression	CD133, CD133 ⁺ M
Oncogenes	C-MYC
Tumorigenic	Yes, tumorigenic in nude mice
Viruses	Adenovirus: Adenovirus expressing CD133 (ALV)
Characteristics	
Culture Medium	DMEM, w: 4.5 g/L D-glucose, w: 4 mM L-glutamine, w: 3.7 g/L NaHCO ₃ , w: 1.0 mM Sodium Pyruvate (Cytion 820300a)
Supplements	10% FBS, 0.05 µg/ml 2-Mercaptoethanol, 5% Penicillin, 10% Streptomycin → 2-Mercaptoethanol (Cytion 820300a)
Dissociation Reagent	Trypsin
Subculturing	Cells are grown in DMEM supplemented with 10% FBS, 0.05 µg/ml 2-Mercaptoethanol, 5% Penicillin, 10% Streptomycin. Cells are passaged by trypsinization.
Seeding density	2 × 10 ⁴ - 4 × 10 ⁵ cells/cm ²
Freeze medium	DMEM supplemented with 10% FBS, 0.05 µg/ml 2-Mercaptoethanol, 5% Penicillin, 10% Streptomycin + 10% DMSO

DT40 | 305249

**Thawing and
Culturing Cells**

1.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
2.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
3.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
4.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
5.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
6.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
7.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.
8.

Remove the vial from the dry ice and immerse it in a water bath at 37°C. Gently swirl the vial to thaw the cells. Once thawed, transfer the cells to a centrifuge tube and centrifuge at 300 x g for 3 minutes. Remove the supernatant and resuspend the cells in 10 ml of DMEM supplemented with 10% FBS. Seed the cells into a T25 flask and incubate for 24 hours.

**Incubation
Atmosphere**

37°C, 5% CO₂, humidified

**Shipping
Conditions**

Shipped on dry ice, stored at -80°C, shipping temperature -78°C

**Storage
Conditions**

Store at -80°C, do not freeze-thaw. Once thawed, store at -150°C for 196 hours.

DT40 | 305249 | DT40 | 305249

Sterility

DT40 cells are tested for sterility using PCR. The results show that the cells are free of contamination. The cells are also tested for mycoplasma contamination and found to be negative.