

**Nalm-6 Cells | 300297**

**Renseignements généraux**

**Description**

The Nalm-6 cell line, derived from the peripheral blood of a patient with B-cell precursor acute lymphoblastic leukemia (ALL), has become a critical tool in leukemia research. The human cell line Nalm 6 encapsulates the biological characteristics of B-cell ALL, providing a unique window into the disease's genomic landscape, including genome instability and DNA repair mechanisms.

The utility of Nalm-6 cells extends to studying the efficacy of available therapeutic targets and existing resistance mechanisms. The cell line's sensitivity to cytotoxic agents and its role in elucidating the homologous recombination (HDR) repair functions are of particular interest, especially concerning the HDR cells' ability to correct DNA damage.

The Nalm6 cell line is a reliable model for studying the complex nature of acute leukemia. It facilitates research into the gene expression profiles involved in glycolysis, lipid and carbohydrates metabolism, and the mTORC1 pathway, highlighting the metabolic reprogramming in leukemia cells. Furthermore, the cell line's application in reverse genetics and whole transcriptome analysis aids in dissecting the intricate molecular networks driving leukemia progression and resistance.

Research utilizing the Nalm-6 cell line, including studies on clonal variants like clone G5 and resistant cell lines such as those with a high HPRT mutation frequency or C9 with resistance index, provides insights into leukemia's heterogeneity. The exploration of leukemia dynamics, especially in the context of glucocorticoid resistance and MSH2 expression, underscores the potential for developing more targeted and effective treatments for ALL.

In summary, the Nalm-6 cell line is a pivotal resource in leukemia research, offering profound insights into B-cell ALL through its applications in studying genomic instability, DNA repair mechanisms, therapeutic target efficacy, resistance mechanisms, and the underlying molecular pathways influencing leukemia's complex biology and heterogeneity.

**Organism** Human

**Tissue** Blood

**Disease** Adult B acute lymphoblastic leukemia

**Synonyms** NALM-6, NALM 6, Nalm 6, NALM6, Nalm6, NALM-6-M1

**Caractéristiques**

**Age** 19 years

**Gender** Male

**Morphology** Round cells

**Nalm-6 Cells | 300297****Cell type** B cell precursor**Growth properties** Suspension**Données réglementaires****Citation** Nalm-6 (Cytion catalog number 300297)**Biosafety level** 1**NCBI\_TaxID** 9606**CellosaurusAccession** CVCL\_0092**Données biomoléculaires****Reverse transcriptase** Negative**Manipulation****Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion article number 820700a)**Supplements** Supplement the medium with 10% FBS**Doubling time** 35 to 40 hours**Subculturing** Maintain cultures by periodically adding or replacing the medium. Initiate cultures with a density of  $5 \times 10^5$  cells/ml and keep the cell concentration within the range of  $3 \times 10^5$  to  $1 \times 10^6$  cells/ml for optimal growth.**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below  $-150^{\circ}\text{C}$  to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a  $37^{\circ}\text{C}$  water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at  $300 \times g$  for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

### Incubation Atmosphere

$37^{\circ}\text{C}$ , 5%  $\text{CO}_2$ , humidified atmosphere.

### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately  $-78^{\circ}\text{C}$  throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about  $-150$  to  $-196^{\circ}\text{C}$ . Storage at  $-80^{\circ}\text{C}$  is acceptable only as a short interim step before transfer to liquid nitrogen.

## Contrôle de la qualité et analyse moléculaire

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**Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.