Product sheet

A204 Cells | 300109

Renseignements généraux

Description	A204 cells are human epithelial cells derived from the muscles of a 1-year-old female patient with rhabdomyosarcoma. With applications in 3D cell culture and tumorigenic properties, A-204 cells provide an opportunity for studying tumour biology and potential therapeutic interventions. Derived from muscle tissue, A-204 cells closely resemble the outer layer of cells found in organs and tissues. The A204 cell line is characterized by its aggressive undifferentiated phenotype, making it a valuable model for investigating the molecular mechanisms of tumorigenesis and metastasis in soft tissue sarcomas. The presence of specific isoenzymes, including AK-1, ES-D, G6PD, GLO-I, Me-2, PGM1, and PGM3, in A-204 cells provides insight into their metabolic characteristics. These isoenzymes may play a role in understanding cellular processes involved in cancer progression and treatment response. These cells exhibit robust growth in vitro and have been used to study cell proliferation, apoptosis, and drug resistance mechanisms. The A204 cell line is also instrumental in the evaluation of new chemotherapeutic agents and in understanding the interaction between rhabdomyosarcoma cells and therapeutic compounds. This cell line serves as an essential tool for cancer researchers aiming to develop more effective treatments for sarcomas and other related malignancies.
Organism	Human
Tissue	Muscle
Disease	Rhabdomyosarcoma
Metastatic site	Primary tumor site (muscle)
Applications	Rhabdomyosarcoma research; pediatric sarcoma biology; muscle differentiation studies; drug sensitivity; preclinical sarcoma models
Synonyms	A-204

Caractéristiques

Age	1 year
Gender	Female
Morphology	Epithelial-like
Cell type	Rhabdomyosarcoma cells

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Growth properties	Adherent

Données réglementaires

Citation	A204 (Cytion catalog number 300109)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1058
GMO Status	No genetic modification; wildtype rhabdomyosarcoma cell line

Données biomoléculaires

Isoenzymes	PGM3, 1, PGM1, 1, ES-D, 1, Me-2, 1, AK-1, 1, GLO-1, 1, G6PD, B
Tumorigenic	In nude mice. Forms small malignant tumors which are conform to embryonal rhabdomyosarcoma.
Ploidy status	Diploid and tetraploid
MSI-status	Stable (MSS)

Manipulation

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Doubling time	26 to 36 hours

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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	1 to 5
Seeding density	0.5 to 1 x 10^4 cells/cm^2
Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, plate the cells at 2 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 to 48 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Contrôle de la qualité et analyse moléculaire

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.