

**NUGC-4 Cells | 305645**

**Renseignements généraux**

**Description**

NUGC-4 is a human gastric cancer cell line established from metastatic paragastric lymph nodes of an adult patient with poorly differentiated adenocarcinoma exhibiting focal signet-ring cell carcinoma features. The cell line was developed from tumor tissues acquired during surgical resection and has been successfully maintained both in vitro and as a transplantable tumor in nude mice. In vitro, NUGC-4 cells grow predominantly as spherical cells, with some free-floating populations, and exhibit epithelial characteristics confirmed via electron microscopy. These include well-developed endoplasmic reticulum, Golgi apparatus, cytoplasmic filaments, and desmosome-like junctions. Notably, cells contain intracytoplasmic microcysts, contributing to their unique morphology.

Chromosomal analysis reveals that NUGC-4 cells possess a near-triploid karyotype with a modal chromosome number ranging from 52 to 54 in vitro and approximately 53 in vivo. The cells display consistent trisomies across several chromosomal groups, though no specific marker chromosomes were identified. Doubling time for NUGC-4 is approximately 29.9 hours, indicating a moderately rapid proliferation rate under standard culture conditions. Among three related gastric cancer lines (NUGC-2, NUGC-3, and NUGC-4), NUGC-4 exhibited the highest in vitro sensitivity to anticancer agents such as mitomycin C and adriamycin, suggesting a heightened responsiveness to certain DNA-damaging chemotherapeutics.

Histologically, xenografts derived from NUGC-4 resemble the parent tumor, maintaining features of a scirrhous carcinoma pattern. The line has been used in drug response profiling and molecular characterization studies as part of large-scale cancer cell line projects. Its combination of clinical origin, histological fidelity, and drug sensitivity profile makes NUGC-4 a relevant model for studying aggressive and chemoresponsive gastric adenocarcinomas with diffuse-type characteristics.

**Organism** Human

**Tissue** Metastatic

**Disease** Gastric signet ring cell adenocarcinoma

**Metastatic site** Paragastric lymph node

**Synonyms** NUGC4, NU-GC-4, Nagoya University-Gastric Cancer-4

**Caractéristiques**

**Age** 35 years

**Gender** Female

**Ethnicity** Japanese

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**Growth properties** Adherent

**Données réglementaires**

**Citation** NUGC-4 (Cytion catalog number 305645)

**Biosafety level** 1

**NCBI\_TaxID** 9606

**CellosaurusAccession** CVCL\_3082

**Données biomoléculaires****Manipulation**

**Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO<sub>3</sub> (Cytion article number 820700a)

**Supplements** Supplement the medium with 10% FBS

**Dissociation Reagent** Accutase

**Doubling time** 29.9 hours

**Seeding density** 1 to 4 x 10<sup>4</sup> cells/cm<sup>2</sup>

**Fluid renewal** 2 to 3 times per week

**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below  $-150^{\circ}\text{C}$  to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a  $37^{\circ}\text{C}$  water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at  $300 \times g$  for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

### Incubation Atmosphere

$37^{\circ}\text{C}$ , 5%  $\text{CO}_2$ , humidified atmosphere.

### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately  $-78^{\circ}\text{C}$  throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about  $-150$  to  $-196^{\circ}\text{C}$ . Storage at  $-80^{\circ}\text{C}$  is acceptable only as a short interim step before transfer to liquid nitrogen.

## Contrôle de la qualité et analyse moléculaire

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### **Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.