Product sheet

SK-N-AS Cells | 305272

Renseignements généraux

Description	The SK-N-AS cell line is derived from a neuroblastoma of a human child and is extensively used in neuro-oncology research. Neuroblastoma is a type of cancer that arises from neural crest cells and predominantly affects children. SK-N-AS cells provide a valuable model for studying the biology and treatment of neuroblastoma, particularly in understanding the molecular mechanisms driving tumor development and progression. This cell line is characterized by its relatively undifferentiated state, which makes it useful for examining the pathways involved in neuronal differentiation and malignancy. SK-N-AS cells exhibit an adherent growth pattern and possess a neuroblastic morphology. They express various markers associated with neural crest cells and neuroblastoma, including neuron-specific enolase (NSE) and chromogranin A. Researchers utilize SK-N-AS cells to investigate the genetic and epigenetic changes associated with neuroblastoma, such as MYCN amplification and ALK mutations. These cells are also employed in high-throughput drug screening and preclinical testing of novel chemotherapeutic agents and targeted therapies. Additionally, SK-N-AS cells are used to study the mechanisms of resistance to conventional therapies and to develop strategies to overcome such resistance. The relevance of SK-N-AS cells in neuroblastoma research underscores their importance in advancing our understanding of this aggressive childhood cancer and in improving therapeutic approaches for affected patients.
Organism	Human
Tissue	Brain
Disease	Neuroblastoma
Metastatic site	Bone marrow
Synonyms	SKN-AS, SKNAS

Caractéristiques

Age	6 years
Gender	Female
Ethnicity	European
Morphology	Epithelial
Cell type	Neuroblast
Growth properties	Adherent

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Données réglementaires

Citation	SK-N-AS (Cytion catalog number 305272)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_1700

Données biomoléculaires

Tumorigenic	Yes, in nude mice
Mutational profile	Mutation: NRAS, p.Gln61Lys (c.181C>A), heterozygous

Manipulation

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS, 1% NEAA
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use 50% basal medium + 40% FBS + 10% DMSO, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Contrôle de la qualité et analyse moléculaire

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.