

CHO-B7H3 Cells | 305417**Información general****Description**

Disclaimer: The prices displayed for cell lines are exclusively for academic/not-for-profit customers. For commercial entities the price is approximately €6,250.

If you represent a commercial entity or are unsure which category applies, please [contact us](#).

The CHO-B7H3 cell line is a stable recombinant CHO (Chinese Hamster Ovary) cell line engineered to express the B7-H3 receptor at a high level, approximately 430,000 molecules per cell. This cell line was developed using innovative landing pad technology, which ensures precise and reproducible integration of the B7-H3 gene at a specific, pre-validated genomic locus. B7-H3, also known as CD276, is a member of the B7 family of immune checkpoint proteins and is overexpressed in various cancers. It plays a critical role in immune evasion by tumor cells and is associated with poor prognosis in cancer patients. This makes B7-H3 a promising target for cancer immunotherapy, particularly in the development of checkpoint inhibitors and antibody-drug conjugates.

The expression of B7-H3 in this cell line was confirmed using flow cytometry with a target-specific antibody, ensuring reliable and consistent receptor density across the cell population.

Organism

Chinese hamster

Tissue

Ovary

Disease

Chinese hamster ovary, non-neoplastic; genetically engineered for B7H3 (CD276) surface expression

Applications

Antibody screening; ADCC/CDC assays; B7H3-targeted therapy development; flow cytometry; drug discovery

Características**Age**

Adult

Gender

Female

Morphology

Epithelial-like

Cell type

Epithelial cells

Growth properties

Adherent/suspension

Datos normativos**Citation**

CHO-B7H3 (Cytion catalog number 305417)

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Biosafety level	1
NCBI_TaxID	10029
CellosaurusAccession	CVCL_A8V5
GMO Status	GMO-S1: This CHO line contains a human B7-H3 expression construct for immune receptor studies. This classification applies only within Germany and may differ elsewhere.

Datos biomoleculares

Receptors expressed	B7H3 (CD276)
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Manejo

Culture Medium	For adherent cultures: DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO ₃ (Cytion article number 820400a) For suspension cultures: CHO Growth Medium A (from InSCREENeX; InSCREENeX catalog number INS-ME-1039)
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Supplements	For adherent cultures: Supplement the medium with 5% FBS. Add Geneticin (G418-Sulfat) to achieve a final concentration of 0.5 mg/mL.
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Dissociation Reagent	For adherent cultures: Trypsin-EDTA
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Doubling time	approx. 14-16 hours
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Subculturing	For routine adherent cell culture: Aspirate the old culture medium from the adherent cells, and wash them with PBS to remove any remaining medium. After aspirating the PBS, add the appropriate volume of Trypsin/EDTA solution based on the culture vessel size (e.g., 1 ml for a T25 flask, 3 ml for a T75 flask) and incubate at room temperature or 37°C for 5-10 minutes, or until the cells detach. Monitor detachment under a microscope, and gently tap the vessel if necessary to release the cells. Once detached, add complete medium to inactivate the Trypsin/EDTA, gently resuspend the cells, and transfer an aliquot of the cell suspension into a new culture vessel containing fresh medium. Place the vessel in an incubator set to 37°C with 5% CO ₂ , and change the medium every 2-3 days.
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Split ratio	1 to 5
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Seeding density	2 to 5 x 10 ⁴ cells/cm ²
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Fluid renewal 2 to 3 times per week

Post-Thaw Recovery

After thawing, split the cells at a ratio of 1:2 to 1:3 in T25 flasks and allow the cells to recover from the freezing process and to adhere (for adherent cultures) for at least 24 hours.

Freeze medium

As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

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Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Control de calidad y análisis molecular

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.