Product sheet

Mv.1.Lu Cells | 305192

General information

Description	The Mv.1.Lu cell line, also known as CCL 64, originates from the lung tissue of a fetal mink (Mustela vison). It is an epithelial-like cell line known for its flat, contact-inhibited growth in monolayer cultures, exhibiting a regular polygonal morphology. This cell line has been widely utilized in virological studies due to its broad permissivity for various mammalian Type C viruses, including murine and feline sarcoma viruses, making it a preferred system for focus formation assays and viral transformation studies. This cell line's ability to support viral replication and transformation has made it an essential tool in understanding viral pathogenesis and host-pathogen interactions. Mv.1.Lu cells are also used in pulmonary physiology research, benefiting from their origin in lung tissue. Studies on their growth characteristics, including their response to various media and culture conditions, have underscored their adaptability and stability in laboratory settings.
Organism	Neovison vison (American mink)
Tissue	Lung
Synonyms	Mv 1 Lu (NBL-7), NBL-7, Mv 1 Lu, MV 1 LU, Mv1.Lu, Mv.1.Lu, MV-1-Lu, Mink, Mink Lung

Characteristics

Age	Near term fetus
Gender	Mixed sex
Morphology	Epithelial
Growth properties	Adherent

Regulatory Data

Citation	Mv.1.Lu (Cytion catalog number 305192)
Biosafety level	1
NCBI_TaxID	452646
CellosaurusAccession	CVCL_0593

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Biomolecular Data

Handling

Culture Medium	EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO3, w: EBSS (Cytion article number 820100a)
Supplements	Supplement the medium with 10% FBS and 1% NEAA
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.