

15P-1 Cells | 305191

General information

Description

15p-1 cells are a mammalian cell line derived from *Mus musculus*, specifically utilized for the study of cellular responses to steroid hormones. Originating from the testicular tissue of mice, these cells exhibit a unique sensitivity to androgens, which makes them particularly valuable in endocrinology and cancer research. The 15p-1 cell line expresses the androgen receptor (AR), enabling the study of androgenic effects on gene expression, cell growth, and differentiation processes.

Characteristically, 15p-1 cells are used to explore the molecular pathways influenced by androgens and their role in diseases such as prostate cancer. They provide a controlled *in vitro* environment to dissect the interactions between androgens and their cellular receptors, facilitating insights into both normal physiological and pathological states. This cell line is also instrumental in screening potential pharmaceuticals targeting androgen-related pathways, contributing to the development of therapeutic strategies.

Organism

Mouse, transgenic

Tissue

Testis

Metastatic site

Primary tumor site (testis)

Applications

Androgen receptor biology; prostate cancer androgen signalling; testicular endocrinology; androgen-responsive gene expression; drug screening for androgen pathway inhibitors

Characteristics

Breed/Subspecies

C57BL/6 x DBA/2

Age

6 months

Gender

Male

Morphology

Epithelial-like

Cell type

Epithelial cells

Growth properties

Adherent

Regulatory Data

Citation

15P-1 (Cytion catalog number 305191)

15P-1 Cells | 305191**Biosafety level** 1**NCBI_TaxID** 10090**CellosaurusAccession** CVCL_6552**GMO Status** GMO-S1: This mouse testis cell line (15P-1) contains the MPyV large T antigen introduced via an MPyV-based vector, supporting transformation and sustained proliferation. The modification is integrated into murine testis-derived cells. This classification applies only within Germany and may differ elsewhere.**Biomolecular Data****Handling****Culture Medium** DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO₃, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)**Supplements** Supplement the medium with 10% FBS**Dissociation Reagent** Accutase**Subculturing** First, remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Split ratio** 1 to 5**Seeding density** 1 to 3×10^4 cells/cm²**Fluid renewal** 2 to 3 times per week**Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

15P-1 Cells | 305191

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.