

Colo-205 Cells | 300380

General information

Description

The COLO-205 cell line is a human colorectal adenocarcinoma cell line first established from the metastatic site of the ascites in a 70-year-old Caucasian male. Characterized by its epithelial cell morphology, this cell line is frequently utilized in biomedical research focused on colorectal cancer, particularly in studies related to cancer biology, drug response, and metastatic mechanisms. COLO-205 cells exhibit a hyperdiploid karyotype and are known to form moderately well-differentiated adenocarcinomas when xenografted into immunodeficient mice.

COLO-205 cells express several key oncogenic and tumor suppressor pathways, making them a valuable model for pharmacological testing and cancer research. They are responsive to tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) making them suitable for apoptosis studies. Furthermore, these cells have been used extensively to investigate the pharmacodynamics of various chemotherapeutic agents, providing insights into the mechanisms of action and resistance in colorectal cancer therapy. Research utilizing the COLO-205 line has contributed significantly to understanding the biological behaviors typical of colorectal adenocarcinomas, including cellular proliferation, differentiation, and interaction with anticancer drugs.

Organism

Human

Tissue

Colon, Dukes' type D

Disease

Colorectal adenocarcinoma

Metastatic site

Ascites

Synonyms

Colo 205, CoLo 205, COLO-205, COLO 205, COLO.205, Colo205, COLO205, Co 205, Colorado 205

Characteristics

Age

70 years

Gender

Male

Morphology

Epithelial-like

Growth properties

Adherent

Regulatory Data

Citation

COLO-205 (Cytion catalog number 300380)

Biosafety level

1

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NCBI_TaxID 9606

CellosaurusAccession CVCL_0218

Biomolecular Data

Protein expression CSAp- (Centriole and Spindle-Associated protein)**Antigen expression** The cells are positive for keratin by immunoperoxidase staining.**Isoenzymes** G6PD, B, PGM1, 1-2, PGM3, 1-2, 6PGD, A, ES-D, 1-2, PEP-D, 1**Tumorigenic** Yes, in nude mice**Reverse transcriptase** Negative**Products** Carcinoembryonic antigen (CEA) 1.5 to 4.1 ng/106 cells/10 days, keratin, interleukin 10 (IL-10, interleukin-10)**Ploidy status** Aneuploid**MSI-status** Stable (MSS)

Handling

Culture Medium RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements** Supplement the medium with 10% FBS**Doubling time** 20 to 25 hours**Subculturing** Collect suspension cells in a 15 ml tube and carefully rinse the adherent cells using PBS without calcium and magnesium (3-5 ml PBS for T25, 5-10ml for T75 cell culture flasks). Add Accutase (1-2ml per T25, 2.5ml per T75 cell culture flask), the cell sheet must be covered completely. Incubate at ambient temperature for 10 minutes, then centrifuge the cells growing in suspension and the adherent cells together. Carefully resuspend the cells and dispense into new flasks which contain fresh medium.**Seeding density** 1×10^4 cells/cm²

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Fluid renewal 2 to 3 times per week

Post-Thaw Recovery After thawing, plate the cells at 5×10^4 cells/cm² and allow the cells to recover from the freezing process and to adhere for at least 24 hours.

Freeze medium As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere 37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

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Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.