

## CFPAC-1 Cells | 305066

## General information

## Description

CFPAC-1 cells, derived from a 26-year-old male with cystic fibrosis and liver metastasis of ductal adenocarcinoma, are a hyperdiploid cell line with notable features for biological research. Their adherence growth property and tumorigenic capability in nude mice make them a practical model for in vitro cancer studies. The cell line's karyotype includes a modal number of 73 chromosomes with several translocations, and importantly, two to three copies of chromosome 7, where the cystic fibrosis gene is located.

These cells express cancer-related antigens and genes like CA19-9, carcinoembryonic antigen (CEA), pancreatic oncofetal antigen (POA), adenocarcinoma associated antigen (ACAA), and epithelial keratins, offering insights into cancer biology. In terms of cystic fibrosis pathology, CFPAC-1 cells demonstrate unique ion transport activities. They do not respond to cAMP agonists, adenylyl cyclase stimulators, or phosphodiesterase inhibitors for chloride ion flux but show increased chloride efflux in response to calcium ionophores.

CFPAC-1 cells carry the common cystic fibrosis mutation - deletion of three nucleotides leading to phenylalanine absence at position 508 in the CFTR gene. Morphologically, they exhibit epithelial features with apical microvilli, tight junctions, and gap junctions, relevant for studying epithelial tissue interactions in both cancer and cystic fibrosis.

**Organism** Human

**Tissue** Pancreas

**Disease** Cystic fibrosis, Pancreatic ductal adenocarcinoma

**Metastatic site** Liver

**Synonyms** CFPac-1, CF PAC-1, CF-PAC1, CF-Pac1, CF Pac1, CFPAC1, CFPac1, CFPAC

## Characteristics

**Age** 26 years

**Gender** Male

**Ethnicity** European

**Morphology** Epithelial

**Growth properties** Adherent

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## Regulatory Data

|                      |                                        |
|----------------------|----------------------------------------|
| Citation             | CFPAC-1 (Cytion catalog number 305066) |
| Biosafety level      | 1                                      |
| NCBI_TaxID           | 9606                                   |
| CellosaurusAccession | CVCL_1119                              |

## Biomolecular Data

|                    |                                                                                                                                                                                             |
|--------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Protein expression | Carcinoembryonic Antigen(Cea), 9Ng/ML, Pancreatic Oncofetal Antigen(Poa), 28Ng/ML, Adenocarcinoma Associated Antigen(Acaa), 5000Ng/ML, Ca 19-9 Antigen, 12000 Units/ML, Epithelial Keratins |
| Antigen expression | CA19-9 antigen, 12000 units/mL, epithelial keratins                                                                                                                                         |
| Tumorigenic        | Yes                                                                                                                                                                                         |

## Handling

|                      |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                         |
|----------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Culture Medium       | IMDM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 25 mM HEPES, w: 1.0 mM Sodium pyruvate, w: 3.024 g/L NaHCO <sub>3</sub> (Cytion article number 820800a)                                                                                                                                                                                                                                                                                                                                                                                                                                                               |
| Supplements          | Supplement the medium with 10% FBS                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                      |
| Dissociation Reagent | Accutase                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                |
| Subculturing         | Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium. |
| Fluid renewal        | 2 to 3 times per week                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                   |
| Freeze medium        | As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.                                                                                                                                                                                                                                                                                                                                 |

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#### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

#### Incubation Atmosphere

37°C, 5% CO<sub>2</sub>, humidified atmosphere.

#### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

#### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

## Quality Control & Molecular Analysis

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#### **Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.