

Caki-2 Cells | 300140**General information****Description**

Caki-2 is a human clear cell renal cell carcinoma (ccRCC) cell line that exhibits epithelial morphology and adheres during in vitro culture conditions. It serves as an essential preclinical model for the investigation of renal cancer mechanisms and therapeutic responses. The Caki-2 line is particularly notable for its resistance to certain chemotherapeutic agents; it displays decreased sensitivity to 5-fluorouracil and the multi-kinase inhibitor sorafenib, which targets VEGFRs 1-3, PDGFR-b, and Raf-1, in comparison to the Caki-1 cell line. This differential sensitivity is significant for studying drug resistance mechanisms and evaluating new therapeutic strategies in renal cell carcinoma.

The genetic background of Caki-2 cells includes a loss-of-function mutation in the von Hippel-Lindau (VHL) tumor suppressor protein, a hallmark of many ccRCCs that leads to the deregulation of hypoxia-inducible factors (HIFs) and contributes to tumorigenesis. The ability of Caki-2 cells to form tumors in immunocompromised mice makes them a valuable tool for in vivo studies of cancer growth and metastasis, providing insights into the tumor environment and potential therapeutic interventions. Their use extends to exploring the role of VHL in cancer progression and testing the efficacy of drugs targeting the HIF pathway and other associated signaling cascades in a controlled experimental setup.

Organism

Human

Tissue

Kidney

Disease

Papillary carcinoma

Synonyms

CAKI-2, CaKi-2, caki-2, CAKI 2, Caki 2, Caki2, CAKI2

Characteristics**Age**

69 years

Gender

Male

Ethnicity

Caucasian

Morphology

Epithelial-like. Ultrastructural features include microvilli and microfilaments. Few mitochondria, lysosomes or lipid droplets. Frequent multilamellar bodies. No virus particles.

Growth properties

Monolayer, adherent

Regulatory Data

Caki-2 Cells | 300140**Citation** Caki-2 (Cytion catalog number 300140)**Biosafety level** 1**NCBI_TaxID** 9606**CellosaurusAccession** CVCL_0235**Biomolecular Data****Isoenzymes** Me-2, 1, PGM3, 1, PGM1, 1, ES-D, 1, AK-1, 1, GLO-1, 1-2, G6PD, B, Phenotype Frequency Product: 0.0511**Tumorigenic** Yes, in nude mice. Forms clear cell carcinoma**Karyotype** (P8) hypopentaploid to hypohexaploid (+A2, +A3, +B, +C, +D, +F, +G, -A) with abnormalities including dicentrics, acrocentric fragments, minutes, breaks, and large subtelocentric markers**Handling****Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements** Supplement the medium with 10% FBS**Dissociation Reagent** Accutase**Subculturing** Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.**Seeding density** 1×10^4 cells/cm² will result in a 90% confluent monolayer in about 4 days**Fluid renewal** 2 to 3 times per week**Post-Thaw Recovery** After thawing, plate the cells at 5×10^4 cells/cm² and allow the cells to recover from the freezing process and to adhere for at least 24 hours.

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Freeze medium

As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

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Quality Control & Molecular Analysis

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.