

**A7r5 Cells | 305198****General information****Description**

Derived from the smooth muscle of the embryonic thoracic aorta in a BDlx rat, the A7r5 cell line is extensively employed in cardiovascular research. These fibroblast-like cells display a unique flat ribbon-like morphology that transitions into parallel arrays of spindle-shaped cells as they differentiate. This distinct structural adaptation facilitates the study of cellular dynamics and morphology under various physiological conditions. During the stationary phase of their growth cycle, A7r5 cells exhibit a significant increase in the activities of myokinase and creatine phosphokinase (CPK), enzymes critical in cellular energy transfer and metabolism.

The synthesis of a specific muscle type CPK isoenzyme upon cessation of cell division in A7r5 cells provides a valuable model for investigating molecular mechanisms underlying muscle development and differentiation. This cell line has been instrumental in exploring the effects of angiotensin II on vascular oxidative stress, offering insights into how this hormone influences cardiovascular physiology. Additionally, A7r5 cells have been used to study the inhibitory effects of phospholipase A2 (PLA2) on lipid droplet formation, further highlighting their utility in cardiovascular research. These applications underscore the A7r5 cell line's versatility and its pivotal role in elucidating critical pathways and potential therapeutic targets in cardiovascular disease studies.

**Organism**

Rat

**Tissue**

Aorta, thoracic, smooth muscle

**Synonyms**

A7R5

**Characteristics****Breed/Subspecies**

BDlx

**Age**

Embryo

**Morphology**

Fibroblast

**Growth properties**

Adherent

**Regulatory Data****Citation**

A7r5 (Cytion catalog number 305198)

**Biosafety level**

1

**NCBI\_TaxID**

10116

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CellosaurusAccession CVCL\_0137

## Biomolecular Data

|                           |                                                             |
|---------------------------|-------------------------------------------------------------|
| <b>Protein expression</b> | Myokinase, Creatine Phosphokinase(Muscle Isoenzyme), Myosin |
|---------------------------|-------------------------------------------------------------|

## Handling

|                       |                                                                                                                                          |
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| <b>Culture Medium</b> | DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO <sub>3</sub> , w: 1.0 mM Sodium pyruvate (Cytion article number 820300a) |
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|                    |                                    |
|--------------------|------------------------------------|
| <b>Supplements</b> | Supplement the medium with 10% FBS |
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|                             |          |
|-----------------------------|----------|
| <b>Dissociation Reagent</b> | Accutase |
|-----------------------------|----------|

|                     |                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                         |
|---------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| <b>Subculturing</b> | Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium. |
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|----------------------|-----------------------|
| <b>Fluid renewal</b> | 2 to 3 times per week |
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|                      |                                                                                                                                                                                                                                                                                         |
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| <b>Freeze medium</b> | As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress. |
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#### Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

#### Incubation Atmosphere

37°C, 5% CO<sub>2</sub>, humidified atmosphere.

#### Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

#### Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

## Quality Control & Molecular Analysis

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#### **Sterility**

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.