Product sheet

CLS-CD-3575 Cells | 400146

General information

Description	CLS-CD-3575 is a human cancer cell line included in curated cell line collections for oncological research. It is derived from a solid tumor of epithelial origin obtained from an adult patient and has been adapted to continuous in vitro culture. The cells grow adherently under standard culture conditions and exhibit morphology consistent with their tissue of origin, forming monolayers with epithelial-like characteristics. As with many established carcinoma cell lines, CLS-CD-3575 demonstrates stable proliferation and suitability for routine passaging. Molecularly, CLS-CD-3575 displays genomic alterations typical of malignant epithelial tumors, including chromosomal imbalances and deregulated signaling pathways associated with proliferation and survival. Depending on the specific tumor origin, expression of lineage-associated cytokeratins and tumor-related markers may be detected. Such features make the line suitable for studies of oncogenic signaling, cell cycle regulation, apoptosis, and drug response profiling in vitro. CLS-CD-3575 is used in experimental settings including cytotoxicity testing, molecular pathway analysis, and evaluation of targeted therapeutic strategies. Its reproducible growth characteristics and compatibility with standard biochemical, molecular biology, and imaging techniques make it a practical model for mechanistic cancer research and preclinical compound screening.
Organism	Mouse
Tissue	Kidney
Disease	Carcinoma
Synonyms	CLS-CD3575

Characteristics

Age	Unspecified
Gender	Unspecified
Growth properties	Adherent

Regulatory Data

Citation	CLS-CD-3575 (Cytion catalog number 400146)
Biosafety level	1

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NCBI_TaxID	10090
CellosaurusAccession	CVCL_5730

Biomolecular Data

Tumorigenic	Yes, in syngeneic mice

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Seeding density	2 to 3 x 10^4 /cm2
Fluid renewal	2 to 3 times per week
Post-Thaw Recovery	After thawing, plate the cells at 5 x 10^4 cells/cm^2 and allow the cells to recover from the freezing process and to adhere for at least 24 hours.
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.