

SK-HEP-1 Cells | 300334

General information

Description The SK-HEP-1 cell line is a cancer cell line derived from a liver adenocarcinoma in a 52-year old Caucasian man. It has been shown to form tumors in immunocompromised mice, produce fibronectin, alpha-1 protease inhibitor, and Interleukin-1. However, there is an alternative hypothesis that the cells are of endothelial origin and not hepatocytes.

Organism Human

Tissue Liver

Disease Adenocarcinoma

Metastatic site Ascites, endothelial cells

Synonyms SK-Hep-1, SK HEP-1, SK HEP 01, SK-Hep1, Sk-Hep1, SK Hep1, SKHEP-1, SKHEP1, SKHep1, SK_HEP1

Characteristics

Age 52 years

Gender Male

Ethnicity Caucasian

Morphology Epithelial-like

Growth properties Adherent

Regulatory Data

Citation SK-HEP-1 (Cytion catalog number 300334)

Biosafety level 1

NCBI_TaxID 9606

CellosaurusAccession CVCL_0525

Biomolecular Data

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Isoenzymes	Me-2, 1-2, PGM3, 1, PGM1, 2, ES-D, 1, AK-1, 1, GLO-1, 1, G6PD, B
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Tumorigenic	Yes, in nude mice, forms large cell carcinoma consistent with hepatoma
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Karyotype	(P11) hyperdiploid to hypotriploid (+A3, +C, +E, +F, +G, -A, -D) with abnormalities including dicentrics, acrocentric fragments, secondary constrictions, pulverizations, and large subtelocentric and submetacentric markers
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Handling

Culture Medium	EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO ₃ , w: EBSS (Cytion article number 820100a)
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Supplements	Supplement the medium with 10% FBS and 1% NEAA
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Dissociation Reagent	Accutase
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Seeding density	1×10^4 cells/cm ²
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Fluid renewal	2 to 3 times per week
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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.