Product sheet

P3X63Ag8 Cells | 305171

General information

Description	This cell ine is derived from the P3K27 cell line (a tissue culture line from the MOPC-21 plasmacytoma in BALB/c mice). The cells are resistant to 8-azaguanine (20 microgram/mL) but sensitive to HAT. Due to a deficiency in 3-ketosteroid reductase activity, the cells have been reported to be cholesterol auxotrophs. This cell line was tested for being not infected by ectromelia virus (mousepox).
Organism	Mouse
Tissue	B Lymphocyte, Phlogocyte, Myeloma
Synonyms	P3x63Ag8, P3-x63-Ag8, P3/x63-Ag8, P3/x63 Ag8, P3/x63/Ag8, P3-x63Ag8, P3x63 Ag8, P3x63 Ag8, P3 x 63Ag8, P3 x 63 Ag8, x63-Ag8, x63-AG8, x63-Ag8, P3x63, x63, GM03571

Characteristics

Breed/Subspecies	BALB/c
Gender	Female
Morphology	Lymphoblast
Growth properties	Suspension

Regulatory Data

Citation	P3x63Ag8 (Cytion catalog number 305171)
Biosafety level	1
NCBI_TaxID	10090
CellosaurusAccession	CVCL_3411

Biomolecular Data

Protein expression	Immunoglobulin, monoclonal antibody

P3X63Ag8 Cells | 305171

Antigen expression	H-2d

Handling

Culture Medium	RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO3 (Cytion article number 820700a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Doubling time	16 to 26 hours
Subculturing	Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1 x 10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.
Seeding density	3 to 5 x 10^4 cells/m^2
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

P3X63Ag8 Cells | 305171

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.