

WERI-Rb-1 Cells | 300632**General information****Description**

The WERI-Rb-1 cell line is derived from a retinoblastoma, a rare malignant tumor of the retina that typically manifests in early childhood. This cell line was established to provide a consistent and replicable model for the study of retinoblastoma biology, offering insights into the genetic, molecular, and cellular mechanisms underlying this form of cancer. WERI-Rb-1 cells are particularly valued in oncological research for their utility in investigating the pathophysiological processes and potential therapeutic targets for retinoblastoma.

WERI-Rb-1 cells exhibit characteristics typical of retinoblastoma, including the expression of neuronal markers and the capability to form cell aggregates resembling Flexner-Wintersteiner rosettes, a hallmark of retinoblastoma histology. These cells have been used extensively to study the role of oncogenes and tumor suppressor genes in cancer development, with a focus on the RB1 gene, mutations of which are pivotal in the etiology of retinoblastoma. Furthermore, WERI-Rb-1 serves as an important tool in the evaluation of chemotherapeutic agents and novel drug delivery systems aimed at improving treatment outcomes for retinoblastoma patients.

Organism

Human

Tissue

Eye

Disease

Retinoblastoma

Applications

3D cell culture

Synonyms

WERI-RB-1, WERI-Rb 1, WERI-Rb1, WERI-RB1, WERI Rb-1, WERIRb1, WERI, Wills Eye Research Institute-Retinoblastoma-1

Characteristics**Age**

1 year

Gender

Female

Morphology

Round cells

Growth properties

Suspension

Regulatory Data**Citation**

WERI-Rb-1 (Cytion catalog number 300632)

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Biosafety level 1**NCBI_TaxID** 9606**CellosaurusAccession** CVCL_1792

Biomolecular Data

Isoenzymes ES-D, 1, G6PD, B, GLO-I, 2, Me-2, 1, PGM1, 1, PGM3, 0**Tumorigenic** Yes, in rabbits**Viruses** EBV -, HBV -, HCV -, HHV-8 -, HIV-1 -, HIV-2 -, HTLV-1/2 -, MLV -, SMRV -**Reverse transcriptase** Negative**Karyotype** Human pseudodiploid karyotype with 3.9% polyploidy - 46(41-48)2n>xx, +6, -10, -10, -14, -22, +3mar, add(3)(q25), add(3)(q25), add(4)(p15), add(5)(q35), i(6q), del(7)(p21), add(9)(q33), der(13)x2, add(16)(q23), add(16)(q23), i(17q), add(19)(q13) - apparently (uniparental?) disomic rearrangement of ch 13 - corresponds to reported karyotype

Handling

Culture Medium RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements** Supplement the medium with 10% FBS and 0.01 mg/mL insulin**Subculturing** Gently homogenize the cell suspension in the flask by pipetting up and down, then take a representative sample to determine the cell density per ml. Dilute the suspension to achieve a cell concentration of 1×10^5 cells/ml with fresh culture medium, and aliquot the adjusted suspension into new flasks for further cultivation.**Freeze medium** As a cryopreservation medium, we use 50% basal medium + 40% FBS + 10% DMSO, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.