

HK-CRISPR-mEGFP-Nup214 Cells | 300671**General information****Description**

The HK-CRISPR-mEGFP-Nup214 cell line is designed to study Nup214, a key nuclear pore complex (NPC) component. Using CRISPR-Cas9, this cell line adds a monomeric enhanced green fluorescent protein (mEGFP) tag to Nup214, allowing real-time visualization of its role in nucleocytoplasmic transport and cell cycle regulation.

This cell line ensures minimal off-target effects and enables advanced imaging techniques to study NPC structure and function. It is valuable for investigating nucleocytoplasmic transport and related diseases, such as certain leukemias, and is useful for screening compounds that affect NPC function.

Organism

Human

Tissue

Endocervix

Disease

Adenocarcinoma

Characteristics**Age**

30 years

Gender

Female

Ethnicity

African American

Morphology

Epithelial-like cells with mosaic stone shape

Growth properties

Adherent

Regulatory Data**Citation**

HK-CRISPR-mEGFP-Nup214 (Cytion catalog number 300671)

Biosafety level

1

NCBI_TaxID

9606

Depositor

The Ellenberg Lab (EMBL)

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GMO Status	GMO-S1: This HeLa Kyoto line contains a CRISPR-engineered mEGFP fusion at the Nup214 locus for studying nucleocytoplasmic transport. This classification applies only within Germany and may differ elsewhere.
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Biomolecular Data

Protein expression	Nup214, mEGFP-tag
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Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO ₃ , w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
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Supplements	Supplement the medium with 10% FBS
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Dissociation Reagent	Accutase
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Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
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Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.
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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.