

SW620-Luc Cells | 305704

General information

Description

SW620-Luc is a bioluminescent derivative of the human SW620 metastatic colon adenocarcinoma cell line, engineered to stably express a firefly luciferase reporter gene. The parental SW620 cell line was derived from a lymph node metastasis of the same patient from whom SW480 was established, allowing paired primary-metastatic comparative studies of colorectal cancer biology. SW620 cells exhibit epithelial-like morphology and share the same key oncogenic mutations as SW480, including KRAS G12V, TP53 R273H and P309S, and APC truncation, but display more mesenchymal features and greater invasive potential, consistent with their metastatic origin. SW620 is widely used to study colorectal cancer metastasis, epithelial-to-mesenchymal transition (EMT), and therapeutic resistance mechanisms.

The stable luciferase integration in SW620-Luc enables sensitive, quantitative bioluminescence imaging (BLI) of tumor burden in xenograft and experimental metastasis models in immunocompromised hosts. The emitted signal correlates with viable tumor cell number, supporting longitudinal monitoring of tumor engraftment, growth kinetics, liver colonization, and lymph node dissemination. SW620-Luc is particularly valuable for studying the metastatic progression of colorectal cancer, evaluating anti-metastatic agents, and performing comparative in vivo studies with the matched primary SW480 line.

SW620-Luc retains the molecular profile and metastatic phenotype of the parental SW620 line, including KRAS and TP53 mutations. The luciferase modification substantially enhances experimental throughput and sensitivity for in vivo pharmacodynamic assessment of treatment efficacy. Researchers should verify luciferase activity, mutational profile, and growth kinetics under their specific experimental conditions prior to large-scale preclinical use.

Organism	Human
Tissue	Metastatic
Disease	Colon adenocarcinoma
Metastatic site	Lymph node
Synonyms	SW620, SW 620, SW.620

Characteristics

Age	51 years
Gender	Male
Ethnicity	Caucasian
Morphology	Epithelial-like

SW620-Luc Cells | 305704

Growth properties	Adherent
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Regulatory Data

Citation	SW620-Luc (Cytion catalog number 305704)
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Biosafety level	1
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NCBI_TaxID	9606
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CellosaurusAccession	CVCL_J268
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GMO Status	GMO-S1: This cell line contains a stably integrated firefly luciferase reporter cassette (Luc2, codon-optimized) introduced via replication-incompetent lentiviral transduction. The resulting polyclonal cell population was maintained under puromycin selection (1–5 µg/mL). S1 containment is required. This classification applies only within Germany and may differ elsewhere.
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Biomolecular Data

Antigen expression	Luc2 (firefly, codon-optimized)
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Tumorigenic	Yes, in athymic nude mice
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Mutational profile	Mutation: p.Gln1338Ter, Homozygous; Mutation: p.Gly12Val, Homozygous; Mutation: p.Arg273His, Heterozygous; Mutation: p.Pro309Ser, Heterozygous
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Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 3.7 g/L NaHCO ₃ , w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
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Supplements	Supplement the medium with 10% FBS
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Dissociation Reagent	Accutase
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SW620-Luc Cells | 305704

Subculturing Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Split ratio 1 to 3

Seeding density 1 to 3×10^4 cells/cm²

Fluid renewal 2 to 3 times per week

Freeze medium As a cryopreservation medium, we use complete growth medium + 10% DMSO for adequate post-thaw viability.

Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 200 x g for 5 minutes, carefully discard the supernatant containing freezing medium.
7. Follow the procedure described under Post-Thaw Recovery

Incubation Atmosphere 37°C, 5% CO₂, humidified atmosphere.

SW620-Luc Cells | 305704

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis