

DOHH-2 Cells | 305537

General information

Description

DOHH-2 is a human diffuse large B-cell lymphoma (DLBCL) cell line that is used to model various aspects of B-cell malignancies, including drug resistance and treatment efficacy. This cell line is particularly valuable for assessing the impact of targeted therapies and combination regimens on tumor growth. Studies have shown that DOHH-2 cells respond to the Bcl-2 inhibitor ABT-263 (navitoclax), which has been tested in combination with various chemotherapeutic agents to enhance treatment efficacy. For example, the addition of ABT-263 to etoposide and vincristine has been found to yield additive effects on cell survival and apoptosis, demonstrating its potential to improve outcomes in combination treatments for DLBCL. In vivo models involving DOHH-2 also show that combining ABT-263 with regimens like CHOP can significantly increase tumor response rates and delay tumor growth, underscoring its utility in preclinical drug testing.

DOHH-2 has also been studied for its response to various targeted therapies beyond traditional chemotherapy. In particular, the triple combination of the CD20 antibody obinutuzumab with the Bcl-2 inhibitor venetoclax and the MDM2 inhibitor idasanutlin demonstrated superior efficacy in reducing tumor size and achieving complete remission in preclinical models. This approach highlights DOHH-2's role in advancing novel therapeutic strategies aimed at achieving better long-term control of DLBCL.

Organism

Human

Tissue

Pleural effusion

Disease

Diffuse large B-cell lymphoma

Synonyms

DOHH2, DoHH-2, DOHH-2

Characteristics

Age

60 years

Gender

Male

Ethnicity

Caucasian

Growth properties

Suspension

Regulatory Data

Citation

DOHH-2 (Cytion catalog number 305537)

Biosafety level

1

DOHH-2 Cells | 305537**NCBI_TaxID** 9606**CellosaurusAccession** CVCL_1179**GMO Status** GMO-S1: This human lymphoma cell line (DOHH-2) contains EBV-derived genome fragments due to EBV transformation but does not release infectious virus. The modification is stably present in diffuse large B-cell lymphoma cells. This classification applies only within Germany and may differ elsewhere.**Biomolecular Data****Viruses** Transformant: Epstein-Barr virus (EBV).**Handling****Culture Medium** RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements** Supplement the medium with 10% heat-inactivated FBS**Subculturing** Gather the suspension cells in a 15 ml tube and gently wash the adherent cells with PBS lacking calcium and magnesium (use 3-5 ml for T25 flasks and 5-10 ml for T75 flasks). Apply Accutase (1-2 ml for T25 flasks, 2.5 ml for T75 flasks) ensuring full coverage of the cell layer. Allow the cells to incubate at room temperature for 10 minutes. Following incubation, combine and centrifuge both the suspension and adherent cells. After centrifugation, carefully resuspend the cell pellet and transfer the cell suspension into new flasks containing fresh medium.**Seeding density** $3-5 \times 10^5/\text{ml}$ **Freeze medium** As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C, 5% CO₂, humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196 °C. Storage at -80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.