

SNU-398 Cells | 305274**General information****Description**

The SNU-398 cell line is derived from a hepatocellular carcinoma (HCC) of a human adult. This cell line is extensively used in liver cancer research to study the molecular mechanisms underlying hepatocarcinogenesis, tumor progression, and the development of therapeutic strategies. Hepatocellular carcinoma is a prevalent and deadly form of liver cancer, and SNU-398 cells provide a relevant model for investigating the genetic and epigenetic changes associated with this disease.

SNU-398 cells exhibit an epithelial morphology and express markers characteristic of liver cancer, such as alpha-fetoprotein (AFP) and cytokeratins. They harbor genetic mutations and alterations typical of HCC, including mutations in the TP53 gene, which is commonly associated with many cancers. Researchers utilize SNU-398 cells to explore various signaling pathways involved in liver cancer, such as the Wnt/ β -catenin, PI3K/Akt, and MAPK pathways. These cells are also employed in drug screening assays to evaluate the efficacy of chemotherapeutic agents and targeted therapies, as well as in studies investigating resistance mechanisms to conventional treatments. The SNU-398 cell line's importance in hepatocellular carcinoma research lies in its ability to model liver cancer biology and to contribute to the development of more effective therapies for liver cancer patients.

Organism

Human

Tissue

Liver

Disease

Adult hepatocellular carcinoma

Synonyms

SNU398, NCI-SNU-398

Characteristics**Age**

42 years

Gender

Male

Ethnicity

Korean

Morphology

Epithelial

Growth properties

Adherent

Regulatory Data**Citation**

SNU-398 (Cytion catalog number 305274)

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Biosafety level

1

NCBI_TaxID

9606

CellosaurusAccession

CVCL_0077

Biomolecular Data**Surface antigens**

Blood Type 0, Rh +

Viruses

Transformant: Hepatitis B virus (HBV)

Mutational profile

Mutation: CTNNB1, p.Ser37Cys (c.110C>G), heterozygous; Mutation: TP53, p.Ser215Ile (c.644G>T), heterozygous

Handling**Culture Medium**RPMI 1640, w: 2.0 mM stable Glutamine, w: 2.0 g/L NaHCO₃ (Cytion article number 820700a)**Supplements**

Supplement the medium with 10% heat-inactivated FBS, 25 mM HEPES

Dissociation Reagent

Accutase

Subculturing

Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Fluid renewal

2 to 3 times per week

Freeze medium

As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.