

A549 Cells | 300114

General information

Description

A549 cells, derived from lung adenocarcinoma tissue, are a primary model used in cancer research, particularly in biomedical laboratories focusing on lung-related cancers. A549 cells are commonly used as an in vitro model for studying lung cancer biology, drug screening, and the effects of toxic compounds.

In toxicology research, A549 cells offer a controlled experimental model that enables scientists to explore the mechanisms underlying toxic effects and cellular responses. By understanding these mechanisms, researchers can better assess the safety of substances and potentially mitigate their harmful effects.

A549 carcinoma cells have been extensively used as an in vitro model to study lung cancer pathogenesis and as an alternative tissue culture model for various pulmonary-related research studies in biomedical laboratories. These cells maintain the characteristics of type II alveolar epithelial cells and are used to examine the epithelial responses to various infections and inflammatory stimuli, including lung inflammation.

Furthermore, the human cell line A549 serves as a valuable tool in the development of specific antibodies targeting lung cancer-related proteins or markers. By exposing these cells to substances of interest, researchers can investigate how they affect cell viability, proliferation, apoptosis, and other cellular processes. This information aids in the identification of potential therapeutic targets and the development of novel treatments for lung cancer.

In summary, A549 carcinoma cells are pivotal in cancer research, especially concerning lung-related cancers, serving as an in vitro model for cancer and toxicology research, developing effective treatments, and drug screening.

Organism Human

Tissue Lung

Disease Carcinoma

Synonyms A 549, A-549, NCI-A549, hA54

Characteristics

Age 58 years

Gender Male

Ethnicity Caucasian

Morphology Epithelial-like

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Growth properties Adherent

Identifiers / Biosafety / Citation

Citation A549 (Cytion catalog number 300114)

Biosafety level 1

Expression / Mutation

Protein expression p53 positive

Isoenzymes G6PD, type B

Reverse transcriptase negative

Karyotype A549 cells have the modal chromosome number n2, with some cells with 64 chromosomes.

Handling

Culture Medium DMEM:Ham's F12, w: 3.1 g/L Glucose, w: 1.6 mM L-Glutamine, w: 15 mM HEPES, w: 1.0 mM Sodium pyruvate, w: 1.2 g/L NaHCO₃ (Cytion article number 820400a)

Medium supplements Supplement the medium with 10% FBS

Passaging solution Accutase

Doubling time 28 hours

Subculturing Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.

Split ratio A ratio of 1:4 is recommended

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Seeding density 1 x 10⁴ cells/cm²

Fluid renewal 2 to 3 times per week

Freezing recovery After thawing, plate the cells at 5 x 10⁴ cells/cm² and allow the cells to recover from the freezing process and to adhere for at least 24 hours.

Freeze medium CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.

STR profile

Amelogenin: x,y
CSF1PO: 10,12
D13S317: 11
D16S539: 11,12
D5S818: 11
D7S820: 8,11
TH01: 8,9,3
TPOX: 8,11
vWA: 14
D3S1358: 16
D21S11: 29
D18S51: 13,17
Penta E: 12,13
Penta D: 9,11
D8S1179: 13
FGA: 20
D1S1656: 16,17
D6S1043: 18
D2S1338: 17
D12S391: 18,23
D19S433: 15,15.2

HLA alleles

A*: 01.01.1900 01:01, 01.01.1900 06:01
B*: 18:01:01, 20:03:01
C*: 12:03:01, 16:01:01
DRB1*: 07:01:01, 11:04:01
DQA1*: 02:01:01, 05:05:01
DQB1*: 02:02:01, 03:01:01
DPB1*: 03:01, 06:01
E: 01:01, 01:03