



General information

Description The HCT-8 line is identical to the HRT-18 cell line. The cells are positive for keratin by immunoperoxidase

staining.

Organism Human

Tissue Rectum

Disease Adenocarcinoma

Synonyms HRT18

Characteristics

Age 67 years

Gender Male

Morphology Epithelial-like

Growth properties

Adherent

Identifiers / Biosafety / Citation

Citation HRT-18 (Cytion catalog number 300230)

Biosafety level 1

Depositor Sobrero

Expression / Mutation

Handling

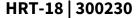
Culture DMEM:Ham's F12, w: 3.1 g/L Glucose, w: 1.6 mM L-Glutamine, w: 15 mM HEPES, w: 1.0 mM Sodium pyruvate, w:

Medium 1.2 g/L NaHCO3 (Cytion article number 820400a)



HRT-18 | 300230

Medium supplements	Supplement the medium with 10% FBS
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:4 to 1:8 is recommended
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)





Handling of cryopreserved cultures

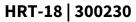
- 1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
- 2. Upon receipt, either store the cryovial immediately at temperatures below -150?C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
- 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37?C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
- 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
- 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
- 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. Optionally, skip centrifugation but remove any remaining freezing medium after 24 hours.
- 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
- 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.





STR profile Amelogenin: x,y

CSF1PO: 12
D13S317: 8,11
D16S539: 12,.13
D5S818: 13
D7S820: 10,12
TH01: 7,9.3
TPOX: 8,11
vWA: 18,19
D3S1358: 17
D21S11: 29,32.2
D18S51: 11,17
Penta E: 7,14
Penta D: 9,14
D8S1179: 15
FGA: 22

HLA alleles A*: 02:01:01, 24:02:01

B*: 08:01:01, 35:01:01 C*: 04:01:01, 07:01:01 DRB1*: 03:01:01, 14:54:01 DQA1*: 01:04:01, 05:01:01 DQB1*: 02:01:01, 05:03:01G DPB1*: 01:01:01, 04:01:01

E: 01:03:02, 01:xx