Product sheet

PC-3M Cells | 305061

General information

Description	The PC-3M cell line is a metastatic variant derived from the human prostate adenocarcinoma PC-3 cell line, originally isolated from a bone metastasis of a prostate cancer patient. PC-3M was established to better model the metastatic potential of prostate cancer. This cell line exhibits enhanced migratory and invasive capabilities compared to its parental counterpart, making it a critical tool in studying the molecular mechanisms of metastasis and evaluating therapeutic interventions targeting metastatic prostate cancer. PC-3M cells have been used in various in vitro and in vivo studies to investigate tumor progression and therapeutic resistance mechanisms. They have shown adaptability to diverse culture conditions and exhibit robust growth both in standard culture and in animal models. Notably, the PC-3M line has been widely applied in xenograft studies, where it demonstrates the ability to form tumors and metastasize efficiently, replicating key characteristics of advanced-stage prostate cancer. This makes it an invaluable model for testing anti-metastatic agents and elucidating pathways that drive metastatic dissemination. In addition to its metastatic properties, PC-3M has been utilized to explore interactions between tumor cells and the microenvironment, including the role of stromal cells and extracellular matrix components in promoting cancer progression. The cell line also expresses biomarkers relevant to prostate cancer, such as prostate-specific antigen (PSA), and is amenable to genomic and proteomic profiling, enabling researchers to investigate molecular pathways and identify potential therapeutic targets.
Organism	Human
Tissue	Prostate
Disease	Prostate carcinoma
Metastatic site	Bone
Synonyms	PC3-M, PC-3/M, PC3M, Pc3M

Characteristics

Age	62 years
Gender	Male
Morphology	Epithelial
Growth properties	Adherent

Regulatory Data

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Citation	PC-3M (Cytion catalog number 305061)
Biosafety level	1
NCBI_TaxID	9606
CellosaurusAccession	CVCL_9555

Biomolecular Data

Handling

Culture Medium	Ham's F12K Medium, w: 2.0 mM L-Glutamine, w: 2.0 mM Sodium pyruvate, w: 2.5 g/L NaHCO3 (Cytion article number 820608a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	2 to 3 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Thawing and Culturing Cells	1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit. 2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required. 3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains. 4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening. 5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently. 6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium. 7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth. 8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.
Incubation Atmosphere	37°C, 5% CO_2, humidified atmosphere.
Shipping Conditions	Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately −78 °C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.
Storage Conditions	For long-term preservation, place vials in vapor-phase liquid nitrogen at about −150 to −196 °C. Storage at −80 °C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

Sterility	Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.