Product sheet

BGM Cells | 302158

General information

Description	BGM (Buffalo Green Monkey) cells are a kidney epithelial cell line derived from the African green monkey, Cercopithecus aethiops. These cells are typically used in virological studies because of their susceptibility to various enteroviruses and other viral pathogens, making them a valuable tool in the study of viral infections and viral-host interactions. Their high permissiveness for viral replication is particularly useful for isolating and propagating enteroviruses, rotaviruses, and adenoviruses, among others. In addition to their use in virology, BGM cells are employed in cytotoxicity testing and vaccine production. They provide a consistent and controlled environment for testing the effects of new drugs and potential vaccines on cellular health and viability. BGM cells are also utilized in genetic studies, particularly in understanding gene expression and signaling pathways involved in viral infection and host response mechanisms. Their robust growth and ease of handling in laboratory settings further contribute to their widespread use in biological research.
Organism	Vervet monkey
Tissue	Kidney
Applications	Isolation of water borne viruses
Synonyms	Buffalo Green Monkey cells, BGMK, Buffalo Green Monkey Kidney cells

Characteristics

Gender	Male
Morphology	Epithelial-like
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	BGM (Cytion catalog number 302158)
Biosafety level	1

Expression / Mutation

Handling

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Culture Medium	EMEM (MEM Eagle), w: 2 mM L-Glutamine, w: 2.2 g/L NaHCO3, w: EBSS (Cytion article number 820100a)
Medium supplements	Supplement the medium with 10% FBS and 1% NEAA
Passaging solution	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Freeze medium	As a cryopreservation medium, use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

Handling of cryopreserved cultures

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at 300 x g for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Quality control / Genetic profile / HLA

Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.