

SW-1116 Cells | 300348**General information**

Description The cell line was isolated from colorectal adenocarcinoma tissue by Leibovitz et al. in 1976. They are positive for keratin by immunoperoxidase staining, but negative for CSAp (CSAp-) and colon antigen 3. The oncogenes c-myc, K-ras, H-ras, myb, sis and fos oncogenes are expressed, N-myc and N-ras expression was not observed. The tumor specific nuclear matrix proteins CC-4, CC-5 and CC-6 are expressed.

Organism Human

Tissue Colon

Disease Adenocarcinoma

Synonyms SW1116, SW 1116

Characteristics

Age 73 years

Gender Male

Ethnicity Caucasian

Morphology Epithelial-like

Growth properties Adherent

Regulatory Data

Citation SW-1116 (Cytion catalog number 300348)

Biosafety level 1

NCBI_TaxID 9606

CellosaurusAccession CVCL_0544

Biomolecular Data

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Protein expression	CEA positive
Isoenzymes	G6PD, B, PGM1, 1, PGM3, 1-2, 6PGD, A, ES-D, 1, PEP-D, 1
Oncogenes	Myc +, myb +, ras +, fos +, sis +, p53 +, abl -, ros -, src -
Tumorigenic	Yes, in nude mice
Reverse transcriptase	Negative
Products	Carcinoembryonic antigen (CEA) 2654 ng/106 cells/10 days, keratin
Mutational profile	SW-1116 cells carry a mutation in codon 12 of Kras gene: GGT(Wt Gly) >GCT(Ala)

Handling

Culture Medium	DMEM:Ham's F12 (1:1), w: 3.1 g/L Glucose, w: 2.5 mM L-Glutamine, w: 15 mM HEPES, w: 0.5 mM Sodium pyruvate, w: 1.2 g/L NaHCO ₃ (Cytion article number 820400a)
Supplements	Supplement the medium with 10% FBS
Dissociation Reagent	Accutase
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Fluid renewal	1 to 2 times per week
Freeze medium	As a cryopreservation medium, we use complete growth medium (including FBS) + 10% DMSO for adequate post-thaw viability, or CM-1 (Cytion catalog number 800100), which includes optimized osmoprotectants and metabolic stabilizers to enhance recovery and reduce cryo-induced stress.

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Thawing and Culturing Cells

1. Confirm that the vial remains deeply frozen upon delivery, as cells are shipped on dry ice to maintain optimal temperatures during transit.
2. Upon receipt, either store the cryovial immediately at temperatures below -150°C to ensure the preservation of cellular integrity, or proceed to step 3 if immediate culturing is required.
3. For immediate culturing, swiftly thaw the vial by immersing it in a 37°C water bath with clean water and an antimicrobial agent, agitating gently for 40-60 seconds until a small ice clump remains.
4. Perform all subsequent steps under sterile conditions in a flow hood, disinfecting the cryovial with 70% ethanol before opening.
5. Carefully open the disinfected vial and transfer the cell suspension into a 15 ml centrifuge tube containing 8 ml of room-temperature culture medium, mixing gently.
6. Centrifuge the mixture at $300 \times g$ for 3 minutes to separate the cells and carefully discard the supernatant containing residual freezing medium.
7. Gently resuspend the cell pellet in 10 ml of fresh culture medium. For adherent cells, divide the suspension between two T25 culture flasks; for suspension cultures, transfer all the medium into one T25 flask to promote effective cell interaction and growth.
8. Adhere to established subculture protocols for continued growth and maintenance of the cell line, ensuring reliable experimental outcomes.

Incubation Atmosphere

37°C , 5% CO_2 , humidified atmosphere.

Shipping Conditions

Cryopreserved cell lines are shipped on dry ice in validated, insulated packaging with sufficient refrigerant to maintain approximately -78°C throughout transit. On receipt, inspect the container immediately and transfer vials without delay to appropriate storage.

Storage Conditions

For long-term preservation, place vials in vapor-phase liquid nitrogen at about -150 to -196°C . Storage at -80°C is acceptable only as a short interim step before transfer to liquid nitrogen.

Quality Control & Molecular Analysis

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Sterility

Mycoplasma contamination is excluded using both PCR-based assays and luminescence-based mycoplasma detection methods.

To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.