Product sheet

SCL II Cells | 300497

General information

Description	The human cell line SCL-II was established from an undifferentiated squamous cell carcinoma of a patient by Boukamp et al. in 1983.
Organism	Human
Tissue	Skin, Face
Disease	Squamous cell carcinoma
Synonyms	SCL-II, SCL-2, SCL2

Characteristics

Age	91 years
Gender	Male
Ethnicity	Caucasian
Growth properties	Adherent

Identifiers / Biosafety / Citation

Citation	SCL II (Cytion catalog number 300497)
Biosafety level	1
Depositor	DKFZ, Heidelberg

Expression / Mutation

Protein expression	P53 (+)
Tumorigenic	Formation of highly differentiated, locally invasive squamous cell carcinoma in Balb/c-nu/nu mice.
Karyotype	Aneuploid (hypodiploid, few metaphases hypotetraploid)

Handling

Culture Medium	DMEM, w: 4.5 g/L Glucose, w: 4 mM L-Glutamine, w: 1.5 g/L NaHCO3, w: 1.0 mM Sodium pyruvate (Cytion article number 820300a)
Medium supplements	Supplement the medium with 10% FBS
Passaging solution	TrypLE Express (Life Technologies)
Doubling time	40 to 50 hours
Subculturing	Remove the old medium from the adherent cells and wash them with PBS that lacks calcium and magnesium. For T25 flasks, use 3-5 ml of PBS, and for T75 flasks, use 5-10 ml. Then, cover the cells completely with Accutase, using 1-2 ml for T25 flasks and 2.5 ml for T75 flasks. Let the cells incubate at room temperature for 8-10 minutes to detach them. After incubation, gently mix the cells with 10 ml of medium to resuspend them, then centrifuge at 300xg for 3 minutes. Discard the supernatant, resuspend the cells in fresh medium, and transfer them into new flasks that already contain fresh medium.
Split ratio	A ratio of 1:5 to 1:10 is recommended
Seeding density	1 x 10^4 cells/cm^2
Fluid renewal	2 times per week
Freeze medium	CM-1 (Cytion catalog number 800100) or CM-ACF (Cytion catalog number 806100)
Handling of cryopreserved cultures	SCL II cells are shipped in a deep-frozen state on dry ice. Upon receipt, confirm that the vial remains frozen. For storage, place the cryovial immediately at temperatures below -150 degrees. If you plan to culture the cells immediately, swiftly thaw the vial by shaking it in a 37 degrees water bath with clean water and an antimicrobial agent for 40-60 seconds. Remove the vial once a small ice clump persists, ensuring it remains cold. Proceed with all subsequent steps under aseptic conditions. In a sterile flow hood, disinfect the cryovial with 70% ethanol. Then, gently open the vial and transfer the cell suspension into a 15 ml centrifuge tube pre-filled with 8 ml of room temperature culture medium. Gently mix the cells. For cell separation, centrifuge at 300 x g for 3 minutes and dispose of the supernatant. Skipping centrifugation is optional, although any residual freezing medium should be removed after 24 hours. Resuspend the pellet gently in 10 ml of fresh culture medium and divide between two T25 culture flasks. Follow the subculture protocol for subsequent steps.

Quality control / Genetic profile / HLA

SCL II Cells | 300497

Sterility	Mycoplasma contamination is rigorously excluded using both PCR-based assays and luminescence-based mycoplasma detection methods. To ensure there is no bacterial, fungal, or yeast contamination, cell cultures are subjected to daily visual inspections.
STR profile	**Amelogenin**: x,y **CSF1PO**: 12 **D13S317**: 8,12 **D16S539**: 10,11 **D5S818**: 9 **D7S820**: 8,12 **TH01**: 8 **TPOX**: 8,11 **vWA**: 15,17 **D3S1358**: 14 **D21S11**: 29 **D18S51**: 17 **Penta E**: 13 **Penta D**: 9,13 **D8S1179**: 12,13 **FGA**: 26
HLA alleles	**A***: 68:02:01 **B***: 07:02:01, 07:02:01